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EFFECTOR PROTEINS OF RAPAMYCIN

Related Applications

5 This application is a continuation-in-part of co-pending patent application Serial
No. 08/384,524, filed February 13, 1995, which is a continuation-in-part of patent
application Serial No. 08/312,023, filed September 26, 1994, now abandoned, which
is a continuation-in-part of patent application Serial No. 08/207,975, filed March 8,
1994, now abandoned.

10

This invention concerns effector proteins of Rapamycin. More particularly, this
invention concerns novel Rapamycin-FKBP12 binding proteins of mammalian origin
for identification, design and synthesis of immunomodulatory, anti-restenosis or anti-
tumor agents.

15

BACKGROUND OF THE INVENTION

Rapamycin is a macrolide antibiotic produced by *Streptomyces hygroscopicus*
which was first characterized via its properties as an antifungal agent. It adversely
20 affects the growth of fungi such as *Candida albicans* and *Microsporum gypseum*.
Rapamycin, its preparation and its antibiotic activity were described in U.S. Patent No.
3,929,992, issued December 30, 1975 to Surendra Sehgal et al. In 1977 Martel, R. R.
et al. reported on immunosuppressive properties of rapamycin against experimental
allergic encephalitis and adjuvant arthritis in the Canadian Journal of Physiological
25 Pharmacology, 55, 48-51 (1977). In 1989, Calne, R. Y. et al. in Lancet, 1989, no. 2,
p. 227 and Morris, R. E. and Meiser, B. M. in Medicinal Science Research, 1989, No.
17, P. 609-10, separately reported on the effectiveness of rapamycin in inhibiting
rejection *in vivo* in allograft transplantation. Numerous articles have followed
describing the immunosuppressive and rejection inhibiting properties of rapamycin, and
30 clinical investigation has begun for the use of rapamycin in inhibiting rejection in
transplantation in man.

Rapamycin alone (U.S. Patent 4,885,171) or in combination with picibanil
(U.S. Patent 4,401,653) has been shown to have antitumor activity. R. R. Martel et al.
35 [Can. J. Physiol. Pharmacol. 55, 48 (1977)] disclosed that rapamycin is effective in
the experimental allergic encephalomyelitis model, a model for multiple sclerosis; in the

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adjuvant arthritis model, a model for rheumatoid arthritis; and effectively inhibited the formation of IgE-like antibodies.

5 The immunosuppressive effects of rapamycin have been disclosed in FASEB 3, 3411 (1989). Cyclosporin A and FK-506, other macrocyclic molecules, also have been shown to be effective as immunosuppressive agents, therefore useful in preventing transplant rejection [FASEB 3, 3411 (1989); FASEB 3, 5256 (1989); R. Y. Calne et al., Lancet 1183 (1978); and U.S. Patent 5,100,899].

10 Rapamycin has also been shown to be useful in preventing or treating systemic lupus erythematosus [U.S. Patent 5,078,999], pulmonary inflammation [U.S. Patent 5,080,899], insulin dependent diabetes mellitus [Fifth Int. Conf. Inflamm. Res. Assoc. 121 (Abstract), (1990)], and smooth muscle cell proliferation and intimal thickening following vascular injury [Morris, R. J. Heart Lung Transplant 11 (pt. 2): 197 (1992)].

15 Mono- and diacylated derivatives of rapamycin (esterified at the 28 and 43 positions) have been shown to be useful as antifungal agents (U.S. Patent 4,316,885) and used to make water soluble prodrugs of rapamycin (U.S. Patent 4,650,803). Recently, the numbering convention for rapamycin has been changed; therefore
20 according to Chemical Abstracts nomenclature, the esters described above would be at the 31- and 42- positions. U.S. Patent 5,118,678 discloses carbamates of rapamycin that are useful as immunosuppressive, anti-inflammatory, antifungal, and antitumor agents. U.S. Patent 5,100,883 discloses fluorinated esters of rapamycin. U.S. Patent 5,118,677 discloses amide esters of rapamycin. U.S. Patent 5,130,307 discloses
25 aminoesters of rapamycin. U.S. Patent 5,117,203 discloses sulfonates and sulfamates of rapamycin. U.S. Patent 5,194,447 discloses sulfonylcarbamates of rapamycin.

U.S. Patent No. 5,100,899 (Calne) discloses methods of inhibiting transplant rejection in mammals using rapamycin and derivatives and prodrugs thereof. Other
30 chemotherapeutic agents listed for use with rapamycin are azathioprine, corticosteroids, cyclosporin (and cyclosporin A), and FK-506, or any combination thereof.

Rapamycin produces immunosuppressive effects by blocking intracellular signal transduction. Rapamycin appears to interfere with a calcium independent

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signalling cascade in T cells and mast cells [Schreiber et al. (1992) *Tetrahedron* 48:2545-2558]. Rapamycin has been shown to bind to certain immunophilins which are members of the FK-506 binding proteins (FKBP) family. In particular, Rapamycin has been shown to bind to the binding proteins, FKBP12, FKBP13, FKBP25 [Galat
5 A. et al., (1992) *Biochemistry* 31(8):2427-2437 and Ferrera A, et al., (1992) *Gene* 113(1):125-127; Armistead and Harding, *Ann. Reports in Med. Chem.* 28:207-215, 1993], and FKBP52 [WO 93/07269]

Rapamycin is able to inhibit mitogen-induced T cell and B cell proliferation as
10 well as proliferation induced by several cytokines, including IL-2, IL-3, IL-4 and IL-6 (reviewed by Sehgal et al., *Med. Research Rev.* 14: 1-22, 1994). It can also inhibit antibody production. Rapamycin has been shown to block the cytokine-induced activation of p70^{S6} kinase which appears to correlate with Rapamycin's ability to decrease protein synthesis accompanying cell cycle progression (Calvo et al., *Proc.*
15 *Natl. Acad. Sci. USA*, 89:7571-7575, 1992; Chung et al., *Cell* 69:1227-1236, 1992; Kuo et al., *Nature* 358:70-73, 1992; Price et al., *Science* 257:973-977, 1992). It also inhibits the activation of cdk2/cyclin E complex (Flanagan et al., *Ann. N.Y. Acad. Sci.*, in press; Flanagan et al, *Mol. Cell biol.*, in press; Flanagan et al., *J. Cell Biochem.* 17A:292, 1993). Rapamycin's effects are not mediated by direct binding to p70^{S6}
20 kinase and cdk2/cyclin E, but by action of the Rapamycin-FKBP complex on upstream component(s) which regulate the activation status of the kinases.

It is generally accepted that the action of immunosuppressive drugs, such as Rapamycin, cyclosporine and FK506, is dependent upon the formation of a complex
25 with their respective intracellular receptor proteins called immunophilins. While the binding of these immunosuppressants with their respective immunophilins inhibits the cis-trans peptidyl prolyl isomerase (PPIase) activity of immunophilins, PPIase inhibition is not sufficient to mediate the immunosuppressive activity (reviewed in Armistead and Harding, *Annual Reports in Med. Chem.* 28:207-215:1993). Two
30 rapamycin analogs which are Diels Alder adducts, one with 4-phenyl-1,2,4-triazoline-3,5-dione, and the second with 4-methyl-1,2,4-triazoline-3,5-dione, bind to FKBP, inhibited its PPIase activity, yet they did not exhibit any detectable immunosuppressive activity. The phenyl-triazolinedione Diels Alder adduct at high molar excess has been shown to competitively inhibit rapamycin's effect on DNA synthesis in mitogen-

stimulated murine thymocyte proliferation (Ocain et al., Biochem. Biophys. Res. Commun. 192:1340, 1993). Recent evidence suggests that the binary immunophilin-drug complex such as cyclophilin-cyclosporin A and FKBP-FK506 gains a new function that enables it to block signal transduction by acting on specific target proteins.

5 The molecular target of both cyclophilin-cyclosporin A and FKBP-FK506 complexes such as has been identified as the Ca^{+2} /calmodulin dependent serine/threonine phosphatase calcineurin (J. Liu et al, Cell 66, 807, 1991; J. Liu et al, Biochemistry 31, 3896, 1992; W.M. Flanagan, et al., Nature 352, 803, 1992; McCaffrey et al., J. Biol. Chem. 268, 3747, 1993; McCaffrey et al., Science 262:750, 1993).

10

Rapamycin's antifungal and immunosuppressive activities are mediated via a complex consisting of Rapamycin, a member of the FK506 binding protein (FKBP) family and at least one additional third protein, called the target of Rapamycin (TOR). The family of FKBP's is reviewed by Armistead and Harding (Annual Reports in Med. Chem, 28:207-215:1993). The relevant FKBP molecule in Rapamycin's antifungal activity has been shown to be FKBP12 (Heitman et al., Science 253:905-909:1993). In mammalian cells, the relevant FKBP's are being investigated. Although two TOR proteins (TOR1 and TOR2) have been identified in yeast (Kunz et al., Cell 73:585-596:1993), the target of Rapamycin in human cells remains elusive. The carboxy terminus of yeast TOR2 has been shown to exhibit 20% identity with two proteins, the p110 subunit of phosphatidylinositol 3-kinase and VPS34, a yeast vacuolar sorting protein also shown to have PI 3K activity. However, J. Blenis et al. (AAI meeting, May, 1993) have reported that Rapamycin-FKBP12 complex does not directly mediate its effects on PDGF stimulated cells via the p110, p85 PI 3K complex.

25

DESCRIPTION OF THE INVENTION

This invention concerns isolated, cloned and expressed proteins which bind to a complex of GST-FKBP12-Rapamycin. These proteins are isolated from membrane preparations of Molt 4 T cell leukemia. The sizes of the four novel proteins are estimated by PAGE migration to be 125 ± 12 kilodaltons (kDa), 148 ± 14 kDa, 208 ± 15 kDa and 210 ± 20 kDa and will be referred to herein and in the claims that follow, as the 125 kDa, 148 kDa, 208 kDa, and 210 kDa, respectively. The four proteins may also be referred to herein as effector proteins.

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The proteins of this invention can be used in screening assays, such as enzyme inhibitor assays and binding assays, to identify endogenous complexes and ligands and novel exogenous compounds (like Rapamycin) which modulate their functions. They can also be used in assays to identify compounds with therapeutic benefit for restenosis, immunomodulation and as antitumor agents. Cloning the proteins of this invention does not only allow the production of large quantities of the proteins, it also provides a basis for the development of related anti-sense therapeutics. The use of cDNA clones to generate anti-sense therapeutics with immunomodulatory activity (for use against transplantation rejection, graft versus host disease, autoimmune diseases such as lupus, myasthenia gravis, multiple sclerosis, rheumatoid arthritis, type I diabetes, and diseases of inflammation such as psoriasis, dermatitis, eczema, seborrhea, inflammatory bowel disease, pulmonary inflammation, asthma, and eye uveitis), antirestenosis and anti-tumor activity is included within the scope of this invention.

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The proteins of the present invention can be isolated from mammalian cells, such as cells of the T cell leukemia cell line, Molt 4 (ATCC 1582, American Type Cell Culture, 12301 Parklawn Drive, Rockville, MD, USA, 20852), the B cell lymphoma, BJAB, or normal human T cells. These mammalian cells can be lysed in a buffer containing protease inhibitors and reducing agent (2-ME), such as hypotonic buffer A (100 mM HEPES, pH 7.5, 20 mM KCl, 1 mM EDTA, 0.4 mM PMSF and 2 mM beta mercaptoethanol (2-ME)). The cell nuclei and unbroken cells are cleared by centrifugation at a temperature which minimizes protein degradation. The membrane fraction of the cells can then be concentrated or pelleted by ultracentrifugation at 100,000 g. Detergent solubilization of the membrane pellet is carried out in a detergent containing buffer such as buffer B (50 mM Tris, pH 7.2, 100 mM NaCl, 20 mM KCl, 0.2 mM PMSF, 1 mM 2-ME, 2 mM CaCl₂, 2 mM MgCl₂, 5 µg/ml aprotinin, leupeptin, pepstatin A and antipain), containing CHAPSO (3-[(3-cholamido-propyl)dimethylammonio]-1-propane sulfonate; 12 mM) or Triton X100 (polyethylene glycol 4-isooctylphenyl ether). The solubilized membrane proteins can then be separated from the debris by 100,000g ultracentrifugation at a temperature which minimizes protein degradation. The supernatant containing solubilized membrane proteins is then preabsorbed with an affinity resin, such as glutathione resin, in the presence of protease inhibitors at a temperature which minimizes protein degradation..

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After centrifugation to remove the resin from the supernatant, the supernatant is then incubated with complexed Rapamycin or Rapamycin analog to FKBP, such as GST-FKBP12--Rapamycin at a temperature which minimizes protein degradation. The mixture of solubilized membrane proteins, incubated with complexed Rapamycin or Rapamycin analog to FKBP, such as GST-FKBP12--Rapamycin, can then be incubated with the affinity resin to bind the complexes of rapamycin or rapamycin analog, FKBP fusion protein and binding proteins at a temperature which minimizes protein degradation. After most non-specific proteins are rinsed away using a detergent containing buffer, such as Buffer C (50 mM Tris, pH 7.2, 100 mM NaCl, 20 mM KCl, 0.2 mM PMSF, 1 mM 2-ME or 10 mM dithiothreitol, 0-5 mM CaCl₂, 0-5 mM MgCl₂, 5 µg/ml aprotinin, leupeptin, pepstatin A and antipain and 0.1% Triton X100) (Polyethylene glycol 4-isooctyl phenyl ether), the proteins are eluted from the resin under denaturing conditions, such as a buffer containing sufficient detergent to dissociate it from resin (e.g. Laemli buffer with or without glycerol or dye, as described by Laemli, Nature 227:680, 1970), or non-denaturing conditions such as a buffer containing an appropriate eluting compound for the affinity column, such as 5 mM glutathione. The proteins can then be separated by size using SDS polyacrylamide gel electrophoresis (SDS-PAGE).

The present invention also includes the genomic DNA sequences for the abovementioned proteins, as well as the cDNA and anti-sense RNA and DNA sequences which correspond to the genes for the abovementioned proteins. The present invention further includes the proteins of other mammalian species which are homologous or equivalent at least in function to the abovementioned proteins, as well as the DNA gene sequences for the homologous or equivalent proteins and the cDNA and anti-sense RNA and DNA sequences which correspond to the genes for the homologous or equivalent proteins.

For the purposes of this disclosure and the claims that follow, equivalents of the proteins of this invention are considered to be proteins, protein fragments and/or truncated forms with substantially similar, but not identical, amino acid sequences to the proteins mentioned above, the equivalents exhibiting rapamycin-FKBP complex binding characteristics and function similar to the proteins mentioned above. Therefore, in this specification and the claims below, references to the 125 kDa, 148

kDa, 208 kDa, and 210 kDa proteins of this invention are also to be understood to indicate and encompass homologous or equivalent proteins, as well as fragmented and/or truncated forms with substantially similar, but not identical, amino acid sequences of the 125 kDa, 148 kDa, 208 kDa, and 210 kDa proteins mentioned above.

5

These proteins or protein homologues or equivalents can be generated by similar isolation procedures from different cell types and/or by recombinant DNA methods and may be modified by techniques including site directed mutagenesis. For example, the genes of this invention can be engineered to express one or all of the
10 proteins as a fusion protein with the fusion partner giving an advantage in isolation (e.g. HIS oligomer, immunoglobulin Fc, glutathione S-transferase, FLAG etc). Mutations or truncations which result in a soluble form can also be generated by site directed mutagenesis and would give advantages in isolation.

15

This invention further includes oligopeptide fragments, truncated forms and protein fragments that retain binding affinity yet have less than the active protein's amino acid sequences. This invention also includes monoclonal and polyclonal antibodies specific for the proteins and their uses. Such uses include methods for screening for novel agents for immunomodulation and/or anti-tumor activity and
20 methods of measuring the parent compound and/or metabolites in biological samples obtained from individuals taking immunosuppressive drugs. The use of the cDNA clone to generate anti-sense therapeutics (Milligan et al, J. Med. Chem. 36:1923-1936, 1993) with immunomodulatory activity (transplantation rejection, graft versus host disease, autoimmune diseases such as lupus, myasthenia gravis, multiple sclerosis,
25 rheumatoid arthritis, type I diabetes, and diseases of inflammation such as psoriasis, dermatitis, eczema, seborrhea, inflammatory bowel disease, pulmonary inflammation, asthma, and eye uveitis), and anti-tumor activity is also included in the present invention.

30

The proteins of this invention can also be made by recombinant DNA techniques familiar to those skilled in the art. That is, the gene of the protein in question can be cloned by obtaining a partial amino acid sequence by digestion of the protein with a protease, such as Lysine C, and isolating the resulting protein fragments by microbore HPLC, followed by fragment sequencing (Matsudaira in A Practical

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Guide to Protein and Peptide Purification for Microsequencing, Academic Press (San Diego, CA, 1989)). The determined sequence can then be used to make oligonucleotide probes which can be used to screen a human cDNA library directly or generate probes by polymerase chain reaction. The library can be generated from
5 human T cells or the cell lines, Molt 4, Jurkat, or other etc. to obtain clones. These clones can be used to identify additional clones containing additional sequences until the protein's full gene, i.e. complete open reading frame, is cloned.

It is known in the art that some proteins can be encoded by an open reading
10 frame which is longer than initially predicted by the size of the protein. These proteins may represent cleavage products of the precursor protein translated from the complete open reading frame (eg. IL-1 beta) or proteins translated using a downstream start codon (eg. Hepatitis B surface antigen). In view of this knowledge, it is understood that the term cDNA as used herein and in the claims below refers to cDNA for the
15 gene's complete open reading frame or any portions thereof which may code for a protein of this invention or the protein's fragments, together or separate, or truncated forms, as previously discussed.

In a complementary strategy, the gene(s) for the proteins of this invention may
20 be identified by interactive yeast cloning techniques using FKBP12:RAPA as a trap for cloning. These strategies can also be combined to quicken the identification of the clones.

The relevant cDNA clone encoding the gene for any of the four proteins can
25 also be expressed in E. coli, yeast, or baculovirus infected cells or mammalian cells using state of the art expression vectors. Isolation can be performed as above or the cDNA can be made as a fusion protein with the fusion partner giving an advantage in isolation (e.g. HIS oligomer, immunoglobulin Fc, glutathione S-transferase, etc). Mutations which result in a soluble form can also be generated by site directed
30 mutagenesis and would give advantages in isolation.

The uses of such cDNA clones include production of recombinant proteins. Further, such recombinant proteins, or the corresponding natural proteins isolated from mammalian cells, or fragments thereof (including peptide oligomers) are useful in

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generation of antibodies to these proteins. Briefly, monoclonal or polyclonal antibodies are induced by immunization with recombinant proteins, or the corresponding natural proteins isolated from mammalian cells, or fragments thereof (including peptide oligomers conjugated to a carrier protein (e.g. keyhole limpet hemocyanin or bovine serum albumin)) of animals using state of the art techniques. The antibodies can be used in the purification process of the natural proteins isolated from mammalian cells or recombinant proteins from E. coli, yeast, or baculovirus infected cells or mammalian cells, or cell products.

10 The uses of such cDNA clones include production of recombinant proteins. Further, such recombinant proteins, or the corresponding natural proteins isolated from mammalian cells, are useful in methods of screening for novel agents such as synthetic compounds, natural products, exogenous or endogenous substrates for immunomodulation and/or antitumor activity. The natural products which may be
15 screened may include, but are not limited to, cell lysates, cell supernatants, plant extracts and the natural broths of fungi or bacteria. As an example of a competitive binding assay, one of these proteins attached to a matrix (either covalently or noncovalently) can be incubated with a buffer containing the compounds, natural products, cell lysates or cell supernatants and a labeled rapamycin:FKBP complex. The
20 ability of the compound, natural products, exogenous or endogenous substrates to competitively inhibit the binding of the complex or specific antibody can be assessed. Examples of methods for labeling the complex include radiolabeling, fluorescent or chemiluminescent tags, fusion proteins with FKBP such as luciferase, and conjugation to enzymes such as horse radish peroxidase, alkaline phosphatase, acetylcholine
25 esterase (ACHE), etc. As an example of an enzymatic assay, the proteins are incubated in the presence or absence of novel agents such as synthetic compounds, natural products, exogenous or endogenous substrates with substrate and the enzymatic activity of the protein can be assessed. Methods of measuring the parent compound and/or metabolites in biological samples obtained from individuals taking
30 immunosuppressive drugs can also be assessed using these proteins.

This invention includes a method for identifying substances which may be useful as immunomodulatory agents or anti-tumor agents, the method utilizing the following steps:

- 10 -

a) combining the substance to be tested with one of the four mammalian proteins (125 kDa, 148kDa, 208 kDa or 210 kDa) of this invention, with the protein being bound to a solid support:

5

b) maintaining the substance to be tested and the protein bound to the solid support of step (a) under conditions appropriate for binding of the substance to be tested with the protein, and

10

c) determining whether binding of the substance to be tested occurred in step (b).

This invention also includes a method for identifying substances which may be useful as immunomodulatory or anti-tumor agents which involves the following steps:

15

a) combining a substance to be tested with one of the mammalian proteins of this invention, the protein being bound to a solid support:

20

b) maintaining the substance to be tested and the protein bound to the solid support of step (a) under conditions appropriate for binding of the substance to be tested with the protein, and

25

c) determining whether the presence of the substance to be tested modulated the activity of the mammalian protein.

This invention further includes a method for detecting, in a biological sample, rapamycin, rapamycin analogs or rapamycin metabolites which, when complexed with a FKBP, bind to one of the four listed proteins of this invention, the method comprising the steps of:

30

a) combining the biological sample with a FKBP to form a first mixture containing, if rapamycin, rapamycin analogs or rapamycin metabolites are present in the biological sample, a rapamycin:FKBP complexes, rapamycin analog:FKBP complexes, or rapamycin metabolite:FKBP complexes;

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b) creating a second mixture by adding the first mixture to one of the proteins of this invention, the protein bound to a solid support ;

5 c) maintaining the second mixture of step (b) under conditions appropriate for binding the rapamycin:FKBP complexes, rapamycin analog:FKBP complexes, or rapamycin metabolite:FKBP complexes, if present, to the protein of this invention; and

10 d) determining whether binding of the rapamycin:FKBP complexes, rapamycin analog:FKBP complexes, or rapamycin metabolite:FKBP complexes and the protein occurred in step (c).

Also included in this invention is the use of the cDNA clones to generate anti-
15 sense therapeutics. This can be accomplished by using state of the art techniques, such as those described in Milligan et al, J. Med. Chem. 36:14:1924-1936. For the purposes of this disclosure and the claims that follow, antisense RNA and DNA are understood to include those RNA and DNA strands derived from a cDNA clone which encodes for one of the four proteins (125 kDa, 148 kDa, 208 kDa or 210 kDa) of the
20 present invention which have a native backbone or those which utilize a modified backbone. Such modifications of the RNA and DNA backbones are described in Milligan et al, J. Med. Chem. 36:14:1924-1936. The antisense compounds created by the state of the art techniques recently described (Milligan et al, J. Med. Chem. 36:14:1924-1936) can be useful in modulating the immune response and thus useful in
25 the treatment or inhibition of transplantation rejection such as kidney, heart, liver, lung, bone marrow, pancreas (islet cells), cornea, small bowel, and skin allografts, and heart valve xenografts; in the treatment or inhibition of autoimmune diseases such as lupus, rheumatoid arthritis, diabetes mellitus, myasthenia gravis, and multiple sclerosis; and diseases of inflammation such as psoriasis, dermatitis, eczema, seborrhea,
30 inflammatory bowel disease, and eye uveitis. The antisense molecules of this invention can have antitumor, antifungal activities, and antiproliferative activities. The compounds of this invention therefore can be also useful in treating solid tumors, adult T-cell leukemia/lymphoma, fungal infections, and hyperproliferative vascular diseases such as restenosis and atherosclerosis. Thus, the present invention also comprises

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methods for treating the abovementioned maladies and conditions in mammals, preferably in humans. The method comprises administering to a mammal in need thereof an effective amount of the relevant antisense therapeutic agent of this invention.

5 When administered for the treatment or inhibition of the above disease states, the antisense molecules of this invention can be administered to a mammal orally, parenterally, intranasally, intrabronchially, transdermally, topically, intravaginally, or rectally.

10 It is contemplated that when the antisense molecules of this invention are used as an immunosuppressive or antiinflammatory agent, they can be administered in conjunction with one or more other immunoregulatory agents. Such other immunoregulatory agents include, but are not limited to azathioprine, corticosteroids, such as prednisone and methylprednisolone, cyclophosphamide, rapamycin,
15 cyclosporin A, FK-506, OKT-3, and ATG. By combining the complexes of this invention with such other drugs or agents for inducing immunosuppression or treating inflammatory conditions, the lesser amounts of each of the agents are required to achieve the desired effect. The basis for such combination therapy was established by Stepkowski whose results showed that the use of a combination of rapamycin and
20 cyclosporin A at subtherapeutic doses significantly prolonged heart allograft survival time. [Transplantation Proc. 23: 507 (1991)].

 Treatment with these antisense compounds will generally be initiated with small dosages less than the optimum dose of the compound. Thereafter the dosage is
25 increased until the optimum effect under the circumstances is reached. Precise dosages will be determined by the administering physician based on experience with the individual subject treated. In general, the antisense compounds of this invention are most desirably administered at a concentration that will afford effective results without causing any harmful or deleterious side effects.

30 In light of the therapeutic value of the abovementioned antisense compounds, this invention also includes pharmaceutical compositions containing the antisense RNA and antisense DNA compounds derived from cDNA clones which encode for the 125 kDa, 148 kDa, 208 kDa and 210 kDa proteins of this invention.

This invention also comprises the following process for isolating the proteins of this invention, as well as the proteins isolated therefrom:

5 A process for isolating proteins from mammalian cells, the process comprising the steps of:

1. The mammalian cells of interest are grown and harvested. As mentioned previously, the cells may be of T cell origin (e.g. T cell lymphomas, 10 leukemias, normal human T cells), B cell origin (e.g. EBV transformed B cells, normal human B cells), mast cells, or other cell sources sensitive to rapamycin. The cells may be processed shortly after harvesting or may be stored frozen, such as in pellets, prior to processing. The cells which are kept frozen may be stored in a dry ice and ethanol bath, stored frozen at -70-80° C until use. This step of growing and harvesting the 15 cells of interest may be seen as the first step of this process or as merely preparatory for the present process.

2. Cells are lysed in a buffer containing a buffering agent (e.g. HEPES, Tris, pH 7.5), low salt (e.g. 10 -50 mM NaCl or KCl), chelating agent 20 (e.g. 1-2 mM EDTA), protease inhibitors (e.g. 0.4 mM PMSF) and a reducing agent (e.g. 2 mM 2-ME or 1-20 mM Dithiothreitol) at a temperature which minimizes protein degradation (e.g. 4 °C). It should be understood that the mammalian cells may be treated in any manner capable of producing cell lysis, including sonic lysis and douncing.

25 3. Unbroken cells and cell nuclei are precleared from lysates by centrifugation at a temperature which minimizes protein degradation (e.g. 4 °C). Centrifugation at, for example, 1600g for 10 minutes has been found sufficient to preclear the unbroken cells and cell nuclei from the lysates. This step, while not 30 mandatory, provides a clearer preparation for the steps that follow.

4. The membrane fraction in the precleared lysate is then concentrated, such as by ultracentrifugation. An example of this concentration would be ultracentrifugation at 100,000 g for 1-1.5 hours.

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5. The membrane proteins (e.g. transmembrane, integral and membrane associated proteins) are then solubilized. This may be accomplished by incubating the pellet of Step 4 in a buffer containing a detergent which solubilizes the proteins without detrimentally denaturing them, a buffering agent (e.g. 20-50 mM Tris or HEPES, pH 7.2), salt (e.g. 100 - 200 mM NaCl + 20 mM KCl), reducing agent (e.g. 1-2 mM 2-ME or 1 - 20 mM dithiothreitol), protease inhibitors (e.g. 0.2 mM PMSF, 5 µg/ml aprotinin, leupeptin, pepstatin A and antipain), divalent cations (e.g. 0-5 mM CaCl₂, 0-5 mM MgCl₂) at a temperature which minimizes protein degradation (e.g. 4° C). Examples of detergents useful in this step are CHAPSO (3-[(3-cholamidopropyl)dimethylammonio]-1-propane sulfonate) or Triton X100 (polyethylene glycol 4-isooctylphenyl ether). After this step, the mixture contains solubilized membrane proteins and non-solubilized cellular debris.

6. The solubilized membrane proteins are separated from the non-solubilized cellular debris, such as by ultracentrifugation (eg 100,000g for 1-1.5 hours) at a temperature which minimizes protein degradation (e.g. 4 °C).

7. The supernatant containing solubilized membrane proteins is incubated with an affinity resin in a buffer containing a buffering agent (e.g. 20-50 mM Tris or HEPES, pH 7.2), salt (e.g. 100 - 200 mM NaCl + 20 mM KCl), reducing agent (e.g. 1-2 mM 2-ME or 10 - 20 mM dithiothreitol), protease inhibitors (e.g. 0.2 mM PMSF, 5 µg/ml aprotinin, leupeptin, pepstatin A and antipain), divalent cations (e.g. 0-5 mM CaCl₂, 0-5 mM MgCl₂) at a temperature and time which allows the absorption of the proteins which bind to affinity resin directly, and minimizes protein degradation (e.g. 4 °C).

8. The resin is then removed from the supernatant by centrifugation at a temperature which minimizes protein degradation (e.g. 4 °C).

9. The supernatant is then incubated with Rapamycin or Rapamycin analog (IC₅₀ in LAF < 500nM) complexed to fusion protein of FKBP12 +protein which enhances the isolation of the desired effector protein and through which the fusion protein binds to an affinity resin or affinity column, such as GST-FKBP12,

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5 Histidine oligomer -FKBP12, biotin-FKBP12, etc., in a buffer containing a buffering agent (e.g. 20-50 mM Tris or HEPES, pH 7.2), salt (e.g. 100 - 200 mM NaCl + 20 mM KCl), reducing agent (e.g. 1-2 mM 2-ME or 1 - 20 mM dithiothreitol), protease inhibitors (e.g. 0.2 mM PMSF, 5 µg/ml aprotinin, leupeptin, pepstatin A and antipain), divalent cations (e.g. 0-5 mM CaCl₂, 0-5 mM MgCl₂) at a temperature and for a time which allows binding of the effector proteins to the fusion FKBP protein:Rapamycin or analog complexes and minimizes protein degradation (e.g. 4 °C and 1-2 hours).

10 10. The mixture of Step 9 containing the effector proteins and fusion FKBP protein:Rapamycin complexes is incubated with an affinity resin at a temperature and for a time which allows binding of the complexes of the effector proteins and fusion FKBP protein:Rapamycin or analog to the affinity resin and minimizes protein degradation (e.g. 4 °C and 0.5-2 hours).

15 11. Most non-specific proteins are rinsed away from the resin using a buffer which dissociates binding of non-specific proteins but not the complex between the desired proteins and RAPA-FKBP, such as a buffer containing a buffering agent (e.g. 20-50 mM Tris or HEPES, pH 7.2), salts (e.g. 100 - 1000 mM NaCl, KCl), reducing agent (e.g. 1-2 mM 2-ME or 10 - 20 mM dithiothreitol), protease inhibitors
20 (e.g. 0.2 mM PMSF, 5 µg/ml aprotinin, leupeptin, pepstatin A and antipain), divalent cations (e.g. 0-5 mM CaCl₂, 0-5 mM MgCl₂) and detergent which dissociates binding of non-specific proteins but not the complex between the four proteins and RAPA-fusion FKBP protein such as Triton X100 (Polyethylene glycol 4-isooctyl phenyl ether).

25 12. The effector proteins and the fusion FKBP protein:Rapamycin complexes are eluted from the resin using an appropriate buffer, such as a buffer containing sufficient detergent to dissociate it from resin (e.g. Laemli buffer with or without glycerol or dye, Laemli, Nature 227:680, 1970), or an appropriate eluting
30 compound for the affinity column, such as glutathione, histidine.

13. The effector proteins can then be separated by size. This may be accomplished in any manner which separates the proteins by size, including, but not

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limited to, polyacrylamide gel electrophoresis and size exclusion column chromatography.

5 It might also be useful to compare the proteins isolated by a control procedure, that is a procedure which substitutes buffer for the rapamycin or rapamycin analog with an IC₅₀ in LAF < 500 nM in step 8, can be used to more easily distinguish proteins which bind to the rapamycin:FKBP complex.

10 The proteins of this invention can also be made by recombinant DNA techniques familiar to those skilled in the art. That is, the gene of the protein in question can be cloned by obtaining a partial amino acid sequence by digestion of the protein with an appropriate endopeptidase, such as Lysine C, and isolating the resulting protein fragments by microbore HPLC, followed by fragment sequencing (Matsudaira in A Practical Guide to Protein and Peptide Purification for
15 Microsequencing, Academic Press, San Diego, CA 1989). The determined sequence can then be used to make oligonucleotide probes which can be used to screen a human cDNA library, such as those for human T cells, Molt 4, Jurkat, etc, to obtain clones. (Sambrook, Fritsch, and Maniatis, Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Press, 1989) These clones can be used to identify additional
20 clones containing additional sequences until the protein's full gene is cloned (Sambrook, Fritsch, and Maniatis, Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Press, 1989). In a complementary strategy, the gene(s) may be identified by interactive yeast cloning techniques using FKBP12:RAPA as a trap for cloning (Chien et al., Proc. Natl. Acad. Sci. 88: 9578-9582, 1991). These strategies
25 can also be combined to quicken the identification of the clones.

The relevant cDNA clone can also be expressed in E.coli, yeast, or baculovirus infected cells or mammalian cells using state of the art expression vectors. Isolation can be performed as above or the cDNA can be made as a fusion protein with the fusion
30 partner giving an advantage in isolation (e.g. HIS oligomer, immunoglobulin Fc, glutathione S-transferase, etc). Mutations which result in a soluble form can also be generated by site directed mutagenesis and would give advantages in isolation.

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Homologs in the mouse, rat, monkey, dog and other mammalian species can be obtained using similar procedures. In addition, upon isolation of the human clone of the proteins, the clone can be used to screen for homologs in other mammalian species. These homologs can also be used to develop binding assays and to set up high through
5 put screening assays for compounds, endogenous ligands, exogenous ligands with immunomodulatory activity.

Compounds, endogenous ligands and exogenous ligands having such immunomodulatory activity would can be useful in modulating the immune response
10 and thus useful in the treatment or inhibition of transplantation rejection such as kidney, heart, liver, lung, bone marrow, pancreas (islet cells), cornea, small bowel, and skin allografts, and heart valve xenografts; in the treatment or inhibition of autoimmune diseases such as lupus, rheumatoid arthritis, diabetes mellitus, myasthenia gravis, and multiple sclerosis; and diseases of inflammation such as psoriasis, dermatitis, eczema,
15 seborrhea, inflammatory bowel disease, and eye uveitis.

The compounds, endogenous ligands and exogenous ligands mentioned above can also have antitumor, antifungal activities, and antiproliferative activities. The compounds of this invention therefore can be also useful in treating solid tumors, adult
20 T-cell leukemia/lymphoma, fungal infections, and hyperproliferative vascular diseases such as restenosis and atherosclerosis.

EXAMPLE 1

25 The proteins of the present invention were isolated utilizing a fusion protein of glutathione S-transferase--FK506 binding protein12 (GST-FKBP). GST-FKBP is produced by a recombinant E. coli containing the plasmid, pGEX-FKBP. The cells were grown, induced with IPTG and the fusion protein was isolated using standard technology described in D.B. Smith and K.S. Johnson, Gene 67, 31, 1988 and K.L.
30 Guan and J.E. Dixon, Anal. Biochem. 192, 262, 1991. The solution containing glutathione and GST-FKBP12 was exchanged 5x using a Centricon 10 filtration unit (Amicon) to remove the glutathione and exchange the buffer.

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Molt 4 cells (1×10^9) were grown in standard media (RPMI 1640 containing 100 U/ml penicillin, 100 ug/ml L-glutamine, 10% FCS). The cells were harvested and rinsed 3x with PBS (50mM phosphate buffer, pH 7.0, 150 mM NaCl), flash frozen in dry-ice ethanol bath and stored at -80°C . On ice, the cells were thawed and lysed using a dounce homogenizer with B pestle in 5 ml of buffer A (10 mM Hepes, pH 7.5, 20 mM KCl, 1 mM EDTA, 0.4 mM PMSF and 2 mM 2-ME). After the debris was cleared by centrifugation at 1600g for 10 min. and the membrane fraction was concentrated by 100,000g centrifugation (1 hour), the 100,000 g pellet was incubated in 3 ml buffer B (50 mM Tris, pH 7.2, 100 mM NaCl, 20 mM KCl, 0.2 mM PMSF, 1 mM 2-ME, 2 mM CaCl_2 , 2 mM MgCl_2 , 5 $\mu\text{g/ml}$ aprotinin, leupeptin, pepstatin A and antipain), containing 12 mM CHAPSO for two hours at 4°C . The solubilized membrane proteins were separated from the debris by a 100,000 g centrifugation. After preabsorption of the supernatant for 3-18 hours with 0.4 ml glutathione sepharose resin swollen in buffer B, the supernatant was incubated with complexed Rapamycin-GST-FKBP12 (preformed by incubation of 660 ug GST-FKBP + 60 ug RAPA in buffer B for 1-2 hours, 4°C) for two hours at 4°C . The supernatant was then incubated for 2 hours at 4°C with 100 ul glutathione resin (1:1 Buffer B). Nonspecific proteins were rinsed 5x with buffer C (buffer B + 0.1% Triton x 100) and the proteins eluted from the resin in Laemli buffer by incubation at 95°C for 3 minutes and microcentrifugation. The proteins were separated by size using a 7% SDS-PAGE followed by silver stain. Four bands corresponding to proteins of molecular weights of 210kDa, 208 kDa, 148 kDa, and 125 kDa were present in higher concentrations in the sample containing RAPA + GST-FKBP12 vs GST-FKBP alone.

The mitogen-stimulated thymocyte proliferation assay called the LAF, can be inhibited by rapamycin or analogs such as demethoxyrapamycin and indicates relative activity of rapamycin analogs in immunosuppression. The same proteins were isolated using GST-FKBP complexed with the immunosuppressive analog, demethoxyrapamycin (Table1). The Diels Alder adducts bound to FKBP12 and inhibited PPIase activity of FKBP12 but did not exhibit detectable immunosuppressive activity and thus do not bind to the target of rapamycin. The use of these two compounds complexed with GST-FKBP12 in the analogous isolation procedure (ie. replacing rapamycin:GST-FKBP12) yielded background levels of the 210kDa proteins (no rapamycin)(Table 1). FK506, is an immunosuppressive compound which binds to

- 19 -

FKBP and and mediates at least some of its effects through the binding of the FK506-FKBP complex with calcineurin. FK506 when complexed with GST-FKBP in an analogous procedure yielded only background levels of the 210 kDa protein (Table 1).

5

TABLE 1
Comparison of Binding of Rapamycin
Analog--FKBP12 complexes to 210 kDa Protein

10	Compound	210 kDa	LAF	PPlase(Ki)
	RAPA	+++	6 nM	0.12nM
	demethoxyrapamycin	+++	58nM	4.4 nM
	Diels Alder adduct (phenyl)	±	>1000nM	12 nM
15	Diels Alder adduct (methyl)	±	>1000nM	12 nM
	FK506	±	3nM*	0.4 nM
	none (FKBP)	±		

(* mechanism of action is different)

20

It is known that rapamycin must bind to a member of the FKBP family in order to mediate its effects. To verify that the proteins of this invention bind to the complex RAPA-GST-FKBP and not individually to rapamycin or FKBP12, a modified isolation procedure was employed. The modification consists of using (1) a rapamycin-42-
25 biotin glycinate ester in place of rapamycin (both exhibit equivalent immunosuppressive activity in the LAF assay), (2) no exogenous FKBP and (3) a streptavidin-conjugated resin in place of glutathione-resin. Only background levels of the 210 kDa protein was isolated using this modified isolation procedure.

30

The 210 kDa protein was isolated using the GST--FKBP12--rapamycin complex from BJAB cells (B cell lymphoma) and normal human T lymphocytes purified by Ficoll-Hypaque and T cell columns.

- 20 -

The results of the partial amino acid composition analysis are set forth in Table 2, below. It should be noted that the percentage of the basic amino acids was not determined.

TABLE 2

5

TABLE 2

Peak Number	Component Name	Retention Time	Peak Area	Response Factor	Peak Height	Concentration No./50ul	
10		9.38					
		11.09					
	1	Asp/Asn	12.06	12.47076	0.02344	0.05142	0.30
	2	Thr	13.05	2.92898	0.00000	0.00985	0.068
15	3	Ser	13.78	6.43968	0.00000	0.01995	0.15
		15.68					
	4	Glu/Gln	16.87	25.47273	0.00000	0.05285	0.59
		Prp	18.24				0.14
20	5	Gly	22.35	21.50384	0.00000	0.04645	0.44
		22.90					
	6	Ala	23.73	16.69160	0.00000	0.03113	0.36
		26.06					
25		28.81					
	7	Val	29.39	4.83196	0.00000	0.00605	0.11
		Met	32.28				
	8	Ile	34.10	3.00560	0.2326	0.00782	0.0699
	9	Leu	35.09	5.73202	0.02331	0.01372	0.1383
	10	nLeu	36.27	20.48232	0.02174	0.04286	0.4453

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TABLE 2 (Cont'd)

	Peak Number	Component Name	Retention Time	Peak Area	Response Factor	Peak Height	Concentration No./50μl
5	11	Tyr	38.33	1.44792	0.02618	0.00226	0.0379
	12	Phe	40.05	1.25017	0.02703	0.00187	0.0338
	13	His	47.79	1.50905	0.02553	0.00580	0.0385
	14		51.80	12.66136	0.00000	0.01960	0.0000
10	15	Lys	53.34	9.90767	0.02283	0.02274	0.2262
Totals				146.53645		0.33436	
15	Not Determined			144.29			

EXAMPLE 2

The 210 kDa (210±20 kDa) protein of this invention was isolated from 4 x 10¹¹ Molt 4 cells using the affinity matrix protocol as described previously. Bound proteins were eluted from the affinity matrix with 1x Laemli buffer without glycerol and dye (0.0625 M Tris-HCl, pH6.8, 2% SDS, 0.37M b-mercaptoethanol) and were concentrated 3 consecutive times by centrifugation using centricon 100 (Amicon, Beverly, MA) at 4 °C the first two times and at 18 °C the third time. The concentrated sample was eluted from the centricon 100 filter by incubating 2 hours at room temperature with an equal volume of 2 x laemli buffer without glycerol and dye the first 2 x and 2 x laemli buffer the third time. The proteins in the sample were separated by PAGE on a 1.5mm thick 7% polyacrylamide gel (38:1). The proteins were transferred to polyvinylidene difluoride, PVDF, (Biorad, Hercules, CA) in 10 x Tris/glycine buffer (Biorad) containing 0.037% SDS at 50 mAmps at 4 °C overnight. The proteins on the PVDF were stained with amido black (Biorad) in 10% ethanol, 2% acetic acid and the appropriate band was excised, rinsed with PBS and water and stored frozen.

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Sequencing

The protein (approx. 3 ug) on the PVDF membrane was digested in situ with trypsin using a modification described by J. Fernandez et al, (Anal.Biochem. 201: 255-64, 1992). Briefly, the PVDF was cut into 1 mm² pieces, prewet, and the protein digested in a 100mM Tris-HCl, pH buffer containing 10% acetonitrile, and 1% reduced triton (CalBiochem) with 0.2ug trypsin at 37 °C for 6 hours followed by addition of 0.2 ug trypsin and incubation overnight. The fragments were eluted from the membrane by sonication and the buffer containing the fragments were separated by microfuge centrifugation. The membranes were backextracted 2x (i.e., 50 ul buffer was added to membranes, sonicated, and centrifuged in a microfuge and solution pooled with the original buffer containing the eluted fragments.) The sample (140-145 ul) was separated by narrow bore high performance liquid chromatography using a Vydac C18 2.1mm x 150 mm reverse phase column on a Hewlett Packard HPLC 1090 with a 40 diode array detector as described previously by W.Lane et al, (J.Protein Chem., 10(2): 151-60, 1991). Multiple fractions were collected and measured for absorption at multiple wavelengths (210, 277 and 292 nm). Optimal fractions were chosen for sequencing based on resolution, symmetry, and ultraviolet absorption and spectra (210 nm, 277 nm and 292 nm).. An aliquot (5%) of the optimal fractions was analyzed for homogeneity and length of fragment by matrix assisted laser desorption time of flight mass spectrometry, MALDE-TOF-MS, on a Finnigan lasermat. Selected optimal fractions were sequenced by automated Edman degradation on an Applied Biosystems 477A protein sequencer using microcartridge and manufacturer's recommended chemistry cycle.

Sequence comparison

Comparison was performed using the Intelligenetics suite (Intelligenetics, CA).

Sequences

Utilizing the methods mentioned above, it was determined that the 210 kDa (210±20 kDa) protein of this invention contains peptide fragments, four of which have amino acid sequences as shown below:

- 23 -

- a) ILLNIEHR;
- B) LIRPYMEPLK;
- 5 c) DXMEAQE; and
- d) QLDHPLPTVHPQVTYAYM(K)

10 Those skilled in the art will recognize the one-letter symbols for the amino acids in question (the definitions for which can also be seen at page 21 of the text *Biochemistry*, Third Edition, W.H. Freeman and Company, © 1988 by Lubert Stryer). Those so skilled will also understand that the X in sequence c) indicates an as yet unidentified amino acid and the parentheses in sequence d) indicates that the amino acid in the position in question is possibly lysine.

15

As mentioned previously, the present invention includes fragmented or truncated forms of the proteins mentioned herein. This includes proteins which have as part or all of their amino acid sequence one or more of the four sequences listed as a)-d), above. For the purposes of the claims, below, the proteins referred to as including
20 one or more of the "internal amino acid sequences" are understood to be any protein which contains one of the sequences listed above, whether the protein is comprised wholly of one or more of the sequences a)-d) or whether one or more of the sequences mentioned above form any portion of the protein. This is understood to include all locations on the protein's amino acid sequence including, but not limited to, those
25 sections of the protein which initiate and terminate the protein's amino acid chain.

These partial amino acid sequences were compared with sequences in the Genbank database. There was identity with the sequence, accession number L34075 (Brown et al., Nature 369, 756-758 (1994)). The cDNA of the SEP gene was cloned
30 as follows: Two micrograms of Molt 4 cDNA (Clontech, Palo Alto, CA) in 1 x PCR buffer (10 mM Tris-HCl, pH 8.3, 50 mM KCl, 1 mM MgCl, 200 µM dATP, 200 µM dTTP, 200 µM dCTP, 200 µM dGTP; Perkin Elmer,) with 1 unit Taq polymerase (Perkin Elmer), was amplified by Polymerase chain reaction (PCR) at 94 C for 30 sec., 66 C for 4 min for 30 cycles, 72 C for 10 min by three separate reactions containing
35 one of the following pairs of oligomers:

- 24 -

CGATCGGTCGACTGCAGCACTTTGGGGATTGTGCTCTC and
GCGGCCGCGAGCTTTCTTCATGCATGACAACAGCCCAGGC; or
GCGGCCGCAAGCTTCAAGTATGCAAGCCTGTGCGGCAAGA and
CGATCGGTCGACACCTTCTGCATCAGAGTCAAGTGGTCA; or
5 GCGGCCGCAAGCTTCCTCAGCTCACATCCTTAGAGCTGCA and
CGATCGGTCGACTTATTACCAGAAAGGGCACCAGCCAATATA.

The oligonucleotides were synthesized and isolated by methods previously
described and known in the art (Chemical and Enzymatic Synthesis of Gene
10 Fragments, ed. by H.G.Gassin and Anne Lang, Verlag Chemie, FLA, 1982). The
resulting PCR products named SEP3, SEP4, and SEP5, respectively, were incubated
at 15°C overnight in buffer containing T4 DNA ligase (1 unit) and 50 ng pcII which
was modified to efficiently ligate PCR products (TA cloning kit, Invitrogen, San
Diego, CA) to yield PCR-pcII ligated products. The PCR-pcII products were
15 transformed into competent *E. coli* INValphaF cells obtained commercially from
Invitrogen. Miniprep DNA was prepared using the Quiagen miniprep kits (Quiagen,
Chatsworth, CA) and the clones containing the appropriate sized PCR product were
identified by restriction enzyme digestion with commercially available HindIII or Sal I,
electrophoresis, and comparison to standards. Sep2 and Sep1 cDNA was made using
20 the TimeSaver cDNA synthesis Kit (Pharmacia, Piscataway, NJ) with the first strand
synthesis reaction containing oligodT (0.13 µg) and 250 pmoles of

CGATCGGTCGACCAGATGAGCACATCATAGCGCTGATGA or
CGATCGGTCGACAAATTCAAAGCTGCCAAGCGTTCGGAG,

25 respectively. Sep2 and Sep1 second strand synthesis was performed using the
TimeSaver cDNA synthesis kit with the addition of 250 pmoles of

GCGGCCGCAAGCTTTGGCTCGAGCAATGGGGCCAGGCA or
30 GCGGCCGCAAGCTTAAGATGCTTGAACCGCACCTGCCG,

respectively. The Sep2 and Sep1 cDNA was then amplified by PCR using

- 25 -

CGATCGGTCGACCAGATGAGCACATCATAGCGCTGATGA and
GCGGCCGCAAGCTTTGGCTCGAGCAATGGGGCCAGGCA or
GCGGCCGCAAGCTTAAGATGCTTGAACCGCACCTGCCG and
CGATCGGTCGACAAATTCAAAGCTGCCAAGCGTTCGGAG,

19.
20
21
22

5. respectively as described above. The Sep2 PCR products were cloned into the TA cloning kit (Invitrogen). The Sep 1 PCR products were digested with Hind III and Sal I, separated from the pCl vector by agarose electrophoresis. The Sep1 (HindIII-SalI) fragment was isolated using the Sephaglas bandprep kit from Pharmacia and cloned into the HindIII and Sal I sites of pUC19 as described (Sambrook et al.,
10 Molecular Cloning Cold Spring Harbor, 1989). Ligation of the isolated Sep2(HindIII, AspI) and Sep3(AspI, SalI) fragments or Sep4(HindIII, AccIII/MroI) and Sep5(AccIII/MroI, Sal I) fragments into pUC18(HindIII, SalI) vector and transformation of competent E. coli INValphaF cells (Invitrogen) was performed by
15 techniques known to those skilled in the art (Sambrook et al., Molecular Cloning Cold Spring Harbor, 1989) to obtain pUC18-Sep 23 and pUC18-Sep45 which contain nucleotides 1468- 5326 and 4964 - 7653, respectively, of the full length clone shown in the attached Sequence No. 1. Ligation of the pUC19-Sep1 (EcoRV, SalI), Sep2345
20 (EcoRV, SalI) fragments and transformation of competent E. coli INValphaF cells (Invitrogen) were performed by techniques known to those skilled in the art (as described by Sambrook et al., Molecular Cloning Cold Spring Harbor, 1989) to obtain the full length clone. The nucleic acid sequence coding for this protein and its amino acid sequence are shown in Sequence No. 1.

25 A fusion protein, called glutathione S transferase-sirolimus effector protein, GST-SEP, was engineered by subcloning the Sep4 and Sep5 fragments into the plasmid, pGEX-KG (Guan, K. and Dixon, J.E. (1991) Anal. Biochem. 192, 262-267) as follows. Briefly, Sep4 was digested with commercially available HindIII restriction enzyme, the restriction site was filled in with the Klenow fragment of DNA polymerase
30 (Gibco), and the DNA was extracted with phenol-chloroform and ethanol precipitated using techniques known by those skilled in the art (Sambrook et al., Molecular Cloning Cold Spring Harbor, 1989). The SEP4 (HindIII-Klenow) was further digested with MroI restriction enzyme, separated from the pCl vector by agarose electrophoresis and isolated as the fragment SEP4-HindIII-Klenow-MroI. Sep5 fragment was prepared by

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digestion with Sall and MroI, separated from the pCII vector by agarose electrophoresis and isolated as the fragment SEP5-Sall-MroI. pGEX-KG (Guan, K. and Dixon, J.E. (1991) Anal. Biochem. 192, 262-267) was digested with Nco I, filled in with the Klenow fragment of DNA polymerase and the DNA was extracted with phenol-chloroform and ethanol precipitated, using techniques of those skilled in the art (Sambrook et al., Molecular Cloning Cold Spring Harbor, 1989). pGEX-KG (NcoI, Klenow) was further digested with Sal I, separated from the undigested vector by agarose electrophoresis and isolated as the vector pGEX-KG-NcoI-Klenow-SalI, using techniques of those skilled in the art. Ligation of the vector, pGEX-KG-NcoI-Klenow-SalI and Sep 4 (HindIII, MroI) and Sep5 (MroI, SalI) fragments and transformation into *E. coli* strain INValphaF cells (Invitrogen) using techniques of those skilled in the art yielded the plasmid, pGEX-Sep45. Other *E. coli* hosts such as BL21 can also be used. The DNA and protein sequence of this fusion protein is shown in Sequence No. 2.

15

Flag sequences and kinase recognition domain of heart muscle kinase can be added at the amino terminal end, by methods known in the art (see Chen et al., *Gene* 1994 Feb. 11; 139 (1): 73-75) within SEP or at the carboxy terminus of SEP, SEP4,5 or other fragments using an oligonucleotide which includes the coding sequence for Asp Tyr Lys Asp Asp Asp Lys. The fusion protein can be isolated by affinity chromatography with anti-flag specific antibodies using the commercially available kits from IBI, New Haven, Conn.

25 Transformed host cells containing sequences of this invention have been deposited with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, Maryland 20852, USA, and have been given the ATCC designations listed below:

	<u>Sequence</u>	<u>ATCC Designation</u>
30	a) pUC19-Sep1(nucleotides 1- 1785 of Sequence No. 1)	ATCC 69756
	b) pUC18-Sep23 (nucleotides 1468- 5326 of Sequence No. 1)	ATCC 69753

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c) pUC18-Sep45 (nucleotides 4964 - 7653 of ATCC 69754
Sequence No. 1)

5 d) pUC19-Sep1-5 (ATCC 69756 1-7653 ATCC 69829
of sequence 1)

e) pGEX-Sep45 plasmids (Sequence 2) ATCC 69755.

10

EXAMPLE 3

The 210 kDa protein of this invention was also isolated by the techniques described in Example 1 utilizing the following rapamycin analogs:

15

a) 42-Deoxy-42-[1-(1,1-dimethylethoxy)-2-oxoethoxy] rapamycin (which is described in U.S. Pat. No. 5,233,036);

b) 42-[O-[(1,1-Dimethylethyl)dimethylsilyl]] rapamycin (described in U.S. Pat. No. 5,120,842);

20

c) Rapamycin 42-ester with N-[1,1-dimethylethoxy)carbonyl]-N-methylglycine (described in U.S. Pat. No. 5,130,307);

d) Rapamycin 42-ester with 5-(1,1-dimethylethoxy)-2-[[1,1-dimethylethoxy)carbonyl]amino]-5-oxopentanoic acid ethyl acetate solvate three quarter hydrate (see U.S. Pat. No. 5,130,307);

25

e) Rapamycin 42-ester with N-[(1,1-dimethylethoxy)carbonyl]glycylglycine hydrate (see U.S. Pat. No. 5,130,307); and

f) Rapamycin 42-ester with N2, N6-bis[(1,1-dimethylethoxy)carbonyl]-L-lysine (see U.S. Pat. No. 5,130,307).

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

- 5
- (i) APPLICANT: Molnar-Kimber, Katherine L.
Failli, Amedeo F.
Caggiano, Thomas J.
10 Nakanishi, Koji
Chen, Yanqiu
- (ii) TITLE OF INVENTION: Effector Proteins of
Rapamycin
- 15 (iii) NUMBER OF SEQUENCES: 2
- (iv) CORRESPONDENCE ADDRESS:
(A) ADDRESSEE: Ronald W. Alice,
American Home Products Corporation
20 (B) STREET: 5 Giralda Farms
(C) CITY: Madison
(D) STATE: New Jersey
(E) COUNTRY: USA
(F) ZIP: 07940-0874
- 25 (v) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Diskette, 3.50 inch, 1.4 Mb storage
30 (B) COMPUTER: Apple Macintosh
(C) OPERATING SYSTEM: Macintosh 7.1
(D) SOFTWARE: Microsoft Word
- 35 (vi) CURRENT APPLICATION DATA:
(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:
- 40 (vii) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: US 08/312,023
(B) FILING DATE: 26-SEPTEMBER-1994
(C) APPLICATION NO: US 08/207,975
45 (E) FILING DATE: 08-MARCH-1994
- (viii) ATTORNEY/AGENT INFORMATION:
(A) NAME: Eck, Steven R.
50 (B) REGISTRATION NUMBER: 36,126
(C) REFERENCE/DOCKET NUMBER: AHP-93167-2-C2
- (ix) TELECOMMUNICATION INFORMATION:
(A) TELEPHONE: (610) 902-2628
55 (B) TELEFAX: (610) 688-0273

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(2) INFORMATION FOR SEQ. ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 7653
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double-stranded
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: sense orientation of double-stranded cDNA to mRNA

(iii) HYPOTHETICAL: no

(iv) ANTISENSE: no

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Molt 4 human T-cell leukemia cells
 (B) STRAIN: ATCC Strain CRL 1582

(xi) SEQUENCE DESCRIPTION: SEQ. ID NO: 1

25	AAG ATG CTT GGA ACC GGA CCT GCC GCC GCC ACC ACC GCT GCC ACC ACA	48
	Met Leu Gly Thr Gly Pro Ala Ala Ala Thr Thr Ala Ala Thr Thr	
	1 5 10 15	
30	TCT AGC AAT GTG AGC GTC CTG CAG CAG TTT GCC AGT GGC CTA AAG AGC	96
	Ser Ser Asn Val Ser Val Leu Gln Gln Phe Ala Ser Gly Leu Lys Ser	
	20 25 30	
35	CGG AAT GAG GAA ACC AGG GCC AAA GCC GCC AAG GAG CTC CAG CAC TAT	144
	Arg Asn Glu Thr Arg Ala Lys Ala Ala Lys Glu Leu Gln His Tyr	
	35 40 45	
40	GTC ACC ATG GAA CTC CGA GAG ATG AGT CAA GAG GAG TCT ACT CGC TTC	192
	Val Thr MET Glu Leu Arg Glu MET Ser Gln Glu Glu Ser Thr Arg Phe	
	50 55 60	
45	TAT GAC CAA CTG AAC CAT CAC ATT TTT GAA TTG GTT TCC AGC TCA GAT	240
	Tyr Asp Gln Leu Asn His His Ile Phe Glu Leu Val Ser Ser Ser Asp	
	65 70 75	
50	GCC AAT GAG AGG AAA GGT GGC ATC TTG GCC ATA GCT AGC CTC ATA GGA	288
	Ala Asn Glu Arg Lys Gly Gly Ile Leu Ala Ile Ala Ser Leu Ile Gly	
	80 85 90 95	
	GTG GAA GGT GGG AAT GCC ACC CGA ATT GGC AGA TTT GCC AAC TAT CTT	336
	Val Glu Gly Gly Asn Ala Thr Arg Ile Gly Arg Phe Ala Asn Tyr Leu	
	100 105 110	
	CGG AAC CTC CTC CCC TCC AAT GAC CCA GTT GTC ATG GAA ATG GCA TCC	384
	Arg Asn Leu Leu Pro Ser Asn Asp Pro Val Val MET Glu MET Ala Ser	
	115 120 125	

- 30 -

	AAG	GCC	ATT	GGC	CGT	CTT	GCC	ATG	GCA	GGG	GAC	ACT	TTT	ACC	GCT	GAG	432
	Lys	Ala	Ile	Gly	Arg	Leu	Ala	MET	Ala	Gly	Asp	Thr	Phe	Thr	Ala	Glu	
			130					135						140			
5	TAC	GTG	GAA	TTT	GAG	GTG	AAG	CGA	GCC	CTG	GAA	TGG	CTG	GGT	GCT	GAC	480
	Tyr	Val	Glu	Phe	Glu	Val	Lys	Arg	Ala	Leu	Glu	Trp	Leu	Gly	Ala	Asp	
		145					150					155					
10	CGC	AAT	GAG	GGC	CGG	AGA	CAT	GCA	GCT	GTC	CTG	GTT	CTC	CGT	GAG	CTG	528
	Arg	Asn	Glu	Gly	Arg	Arg	His	Ala	Ala	Val	Leu	Val	Leu	Arg	Glu	Leu	
	160					165					170					175	
15	GCC	ATC	AGC	GTC	CCT	ACC	TTC	TTC	TTC	CAG	CAA	GTG	CAA	CCC	TTC	TTT	576
	Ala	Ile	Ser	Val	Pro	Thr	Phe	Phe	Phe	Gln	Gln	Val	Gln	Pro	Phe	Phe	
				180						185					190		
20	GAC	AAC	ATT	TTT	GTG	GCC	GTG	TGG	GAC	CCC	AAA	CAG	GCC	ATC	CGT	GAG	624
	Asp	Asn	Ile	Phe	Val	Ala	Val	Trp	Asp	Pro	Lys	Gln	Ala	Ile	Arg	Glu	
			195					200						205			
25	GGA	GCT	GTA	GCC	GCC	CTT	CGT	GCC	TGT	CTG	ATT	CTC	ACA	ACC	CAG	CGT	672
	Gly	Ala	Val	Ala	Ala	Leu	Arg	Ala	Cys	Leu	Ile	Leu	Thr	Thr	Gln	Arg	
			210				215						220				
30	GAG	CCG	AAG	GAG	ATG	CAG	AAG	CCT	CAG	TGG	TAC	AGG	CAC	ACA	TTT	GAA	720
	Glu	Pro	Lys	Glu	MET	Gln	Lys	Pro	Gln	Trp	Tyr	Arg	His	Thr	Phe	Glu	
		225					230					235					
35	GAA	GCA	GAG	AAG	GGA	TTT	GAT	GAG	ACC	TTG	GCC	AAA	GAG	AAG	GGC	ATG	768
	Glu	Ala	Glu	Lys	Gly	Phe	Asp	Glu	Thr	Leu	Ala	Lys	Glu	Lys	Gly	MET	
	240					245				250						255	
40	AAT	CGG	GAT	GAT	CGG	ATC	CAT	GGA	GCC	TTG	TTG	ATC	CTT	AAC	GAG	CTG	816
	Asn	Arg	Asp	Asp	Arg	Ile	His	Gly	Ala	Leu	Leu	Ile	Leu	Asn	Glu	Leu	
					260					265					270		
45	GTC	CGA	ATC	AGC	AGC	ATG	GAG	GGA	GAG	CGT	CTG	AGA	GAA	GAA	ATG	GAA	864
	Val	Arg	Ile	Ser	Ser	MET	Glu	Gly	Glu	Arg	Leu	Arg	Glu	Glu	MET	Glu	
				275					280					285			
50	GAA	ATC	ACA	CAG	CAG	CAG	CTG	GTA	CAC	GAC	AAG	TAC	TGC	AAA	GAT	CTC	912
	Glu	Ile	Thr	Gln	Gln	Gln	Leu	Val	His	Asp	Lys	Tyr	Cys	Lys	Asp	Leu	
			290				295						300				
55	ATG	GGC	TTC	GGA	ACA	AAA	CCT	CGT	CAC	ATT	ACC	CCC	TTC	ACC	AGT	TTC	960
	MET	Gly	Phe	Gly	Thr	Lys	Pro	Arg	His	Ile	Thr	Pro	Phe	Thr	Ser	Phe	
		305					310					315					
60	CAG	GCT	GTA	CAG	CCC	CAG	CAG	TCA	AAT	GCC	TTG	GTG	GGG	CTG	CTG	GGG	1008
	Gln	Ala	Val	Gln	Pro	Gln	Gln	Ser	Asn	Ala	Leu	Val	Gly	Leu	Leu	Gly	
	320					325					330					335	

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	TAC	AGC	TCT	CAC	CAA	GGC	CTC	ATG	GGA	TTT	GGG	ACC	TCC	CCC	AGT	CCA	1056
	Tyr	Ser	Ser	His	Gln	Gly	Leu	MET	Gly	Phe	Gly	Thr	Ser	Pro	Ser	Pro	
					340					345					350		
5	GCT	AAG	TCC	ACC	CTG	GTG	GAG	AGC	CGG	TGT	TGC	AGA	GAC	TTG	ATG	GAG	1104
	Ala	Lys	Ser	Thr	Leu	Val	Glu	Ser	Arg	Cys	Cys	Arg	Asp	Leu	MET	Glu	
				355					360					365			
10	GAG	AAA	TTT	GAT	CAG	GTG	TGC	CAG	TGG	GTG	CTG	AAA	TGC	AGG	AAT	AGC	1152
	Glu	Lys	Phe	Asp	Gln	Val	Cys	Gln	Trp	Val	Leu	Lys	Cys	Arg	Asn	Ser	
			370					375					380				
15	AAG	AAC	TCG	CTG	ATC	CAA	ATG	ACA	ATC	CTT	AAT	TTG	TTG	CCC	CGC	TTG	1200
	Lys	Asn	Ser	Leu	Ile	Gln	MET	Thr	Ile	Leu	Asn	Leu	Leu	Pro	Arg	Leu	
		385					390					395					
20	GCT	GCA	TTC	CGA	CCT	TCT	GCC	TTC	ACA	GAT	ACC	CAG	TAT	CTC	CAA	GAT	1248
	Ala	Ala	Phe	Arg	Pro	Ser	Ala	Phe	Thr	Asp	Thr	Gln	Tyr	Leu	Gln	Asp	
	400					405					410				415		
25	ACC	ATG	AAC	CAT	GCC	CTA	AGC	TGT	GTC	AAG	AAG	GAG	AAG	GAA	CGT	ACA	1296
	Thr	MET	Asn	His	Ala	Leu	Ser	Cys	Val	Lys	Lys	Glu	Lys	Glu	Arg	Thr	
				420					425					430			
30	GCG	GCC	TTC	CAA	GCC	CTG	GGG	CTA	CTT	TCT	GTG	GCT	GTG	AGG	TCT	GAG	1344
	Ala	Ala	Phe	Gln	Ala	Leu	Gly	Leu	Leu	Ser	Val	Ala	Val	Arg	Ser	Glu	
				435				440						445			
35	TTT	AAG	GTC	TAT	TTG	CCT	CGC	GTG	CTG	GAC	ATC	ATC	CGA	GCG	GCC	CTG	1392
	Phe	Lys	Val	Tyr	Leu	Pro	Arg	Val	Leu	Asp	Ile	Ile	Arg	Ala	Ala	Leu	
		450					455						460				
40	CCC	CCA	AAG	GAC	TTC	GCC	CAT	AAG	AGG	CAG	AAG	GCA	ATG	CAG	GTG	GAC	1440
	Pro	Pro	Lys	Asp	Phe	Ala	His	Lys	Arg	Gln	Lys	Ala	MET	Gln	Val	Asp	
		465				470						475					
45	GCC	ACA	GTC	TTC	ACT	TGC	ATC	AGC	ATG	CTG	GCT	CGA	GCA	ATG	GGG	CCA	1488
	Ala	Thr	Val	Phe	Thr	Cys	Ile	Ser	MET	Leu	Ala	Arg	Ala	MET	Gly	Pro	
	480					485				490					495		
50	GGC	ATC	CAG	CAG	GAT	ATC	AAG	GAG	CTG	CTG	GAG	CCC	ATG	CTG	GCA	GTG	1536
	Gly	Ile	Gln	Gln	Asp	Ile	Lys	Glu	Leu	Leu	Glu	Pro	MET	Leu	Ala	Val	
				500					505					510			
55	GGA	CTA	AGC	CCT	GCC	CTC	ACT	GCA	GTG	CTC	TAC	GAC	CTG	AGC	CGT	CAG	1584
	Gly	Leu	Ser	Pro	Ala	Leu	Thr	Ala	Val	Leu	Tyr	Asp	Leu	Ser	Arg	Gln	
				515					520					525			
60	ATT	CCA	CAG	CTA	AAG	AAG	GAC	ATT	CAA	GAT	GGG	CTA	CTG	AAA	ATG	CTG	1632
	Ile	Pro	Gln	Leu	Lys	Lys	Asp	Ile	Gln	Asp	Gly	Leu	Leu	Lys	MET	Leu	
			530					535					540				

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	TCC	CTG	GTC	CTT	ATG	CAC	AAA	CCC	CTT	CGC	CAC	CCA	GGC	ATG	CCC	AAG	1680
	Ser	Leu	Val	Leu	MET	His	Lys	Pro	Leu	Arg	His	Pro	Gly	MET	Pro	Lys	
		545					550					555					
5																	
	GGC	CTG	GCC	CAT	CAG	CTG	GCC	TCT	CCT	GGC	CTC	ACG	ACC	CTC	CCT	GAG	1728
	Gly	Leu	Ala	His	Gln	Leu	Ala	Ser	Pro	Gly	Leu	Thr	Thr	Leu	Pro	Glu	
	560					565					570					575	
10																	
	GCC	AGC	GAT	GTG	GGC	AGC	ATC	ACT	CTT	GCC	CTC	CGA	ACG	CTT	GGC	AGC	1776
	Ala	Ser	Asp	Val	Gly	Ser	Ile	Thr	Leu	Ala	Leu	Arg	Thr	Leu	Gly	Ser	
					580					585					590		
15																	
	TTT	GAA	TTT	GAA	GGC	CAC	TCT	CTG	ACC	CAA	TTT	GTT	CGC	CAC	TGT	GCG	1824
	Phe	Glu	Phe	Glu	Gly	His	Ser	Leu	Thr	Gln	Phe	Val	Arg	His	Cys	Ala	
				595					600					605			
20																	
	GAT	CAT	TTC	CTG	AAC	AGT	GAG	CAC	AAG	GAG	ATC	CGC	ATG	GAG	GCT	GCC	1872
	Asp	His	Phe	Leu	Asn	Ser	Glu	His	Lys	Glu	Ile	Arg	MET	Glu	Ala	Ala	
			610					615					620				
25																	
	CGC	ACC	TGC	TCC	CGC	CTG	CTC	ACA	CCC	TCC	ATC	CAC	CTC	ATC	AGT	GGC	1920
	Arg	Thr	Cys	Ser	Arg	Leu	Leu	Thr	Pro	Ser	Ile	His	Leu	Ile	Ser	Gly	
		625					630					635					
30																	
	CAT	GCT	CAT	GTG	GTT	AGC	CAG	ACC	GCA	GTG	CAA	GTG	GTG	GCA	GAT	GTG	1968
	His	Ala	His	Val	Val	Ser	Gln	Thr	Ala	Val	Gln	Val	Val	Ala	Asp	Val	
	640					645					650					655	
35																	
	CTT	AGC	AAA	CTG	CTC	GTA	GTT	GGG	ATA	ACA	GAT	CCT	GAC	CCT	GAC	ATT	2016
	Leu	Ser	Lys	Leu	Leu	Val	Val	Gly	Ile	Thr	Asp	Pro	Asp	Pro	Asp	Ile	
					660					665					670		
40																	
	CGC	TAC	TGT	GTC	TTG	GCG	TCC	CTG	GAC	GAG	CGC	TTT	GAT	GCA	CAC	CTG	2064
	Arg	Tyr	Cys	Val	Leu	Ala	Ser	Leu	Asp	Glu	Arg	Phe	Asp	Ala	His	Leu	
				675					680					685			
45																	
	GCC	CAG	GCG	GAG	AAC	TTG	CAG	GCC	TTG	TTT	GTG	GCT	CTG	AAT	GAC	CAG	2112
	Ala	Gln	Ala	Glu	Asn	Leu	Gln	Ala	Leu	Phe	Val	Ala	Leu	Asn	Asp	Gln	
			690					695					700				
50																	
	GTG	TTT	GAG	ATC	CGG	GAG	CTG	GCC	ATC	TGC	ACT	GTG	GGC	CGA	CTC	AGT	2160
	Val	Phe	Glu	Ile	Arg	Glu	Leu	Ala	Ile	Cys	Thr	Val	Gly	Arg	Leu	Ser	
		705					710					715					
55																	
	AGC	ATG	AAC	CCT	GCC	TTT	GTC	ATG	CCT	TTC	CTG	CGC	AAG	ATG	CTC	ATC	2208
	Ser	MET	Asn	Pro	Ala	Phe	Val	MET	Pro	Phe	Leu	Arg	Lys	MET	Leu	Ile	
		720				725					730					735	
60																	
	CAG	ATT	TTG	ACA	GAG	TTG	GAG	CAC	AGT	GGG	ATT	GGA	AGA	ATC	AAA	GAG	2256
	Gln	Ile	Leu	Thr	Glu	Leu	Glu	His	Ser	Gly	Ile	Gly	Arg	Ile	Lys	Glu	
					740					745					750		

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	CAG	AGT	GCC	CGC	ATG	CTG	GGG	CAC	CTG	GTC	TCC	AAT	GCC	CCC	CGA	CTC	2304
	Gln	Ser	Ala	Arg	MET	Leu	Gly	His	Leu	Val	Ser	Asn	Ala	Pro	Arg	Leu	
				755					760					765			
5	ATC	CGC	CCC	TAC	ATG	GAG	CCT	ATT	CTG	AAG	GCA	TTA	ATT	TTG	AAA	CTG	2352
	Ile	Arg	Pro	Tyr	MET	Glu	Pro	Ile	Leu	Lys	Ala	Leu	Ile	Leu	Lys	Leu	
			770					775					780				
10	AAA	GAT	CCA	GAC	CCT	GAT	CCA	AAC	CCA	GGT	GTG	ATC	AAT	AAT	GTC	CTG	2400
	Lys	Asp	Pro	Asp	Pro	Asp	Pro	Asn	Pro	Gly	Val	Ile	Asn	Asn	Val	Leu	
		785					790					795					
15	GCA	ACA	ATA	GGA	GAA	TTG	GCA	CAG	GTT	AGT	GGC	CTG	GAA	ATG	AGG	AAA	2448
	Ala	Thr	Ile	Gly	Glu	Leu	Ala	Gln	Val	Ser	Gly	Leu	Glu	MET	Arg	Lys	
	800					805					810					815	
20	TGG	GTT	GAT	GAA	CTT	TTT	ATT	ATC	ATC	ATG	GAC	ATG	CTC	CAG	GAT	TCC	2496
	Trp	Val	Asp	Glu	Leu	Phe	Ile	Ile	Ile	MET	Asp	MET	Leu	Gln	Asp	Ser	
					820					825					830		
25	TCT	TTG	TTG	GCC	AAA	AGG	CAG	GTG	GCT	CTG	TGG	ACC	CTG	GGA	CAG	TTG	2544
	Ser	Leu	Leu	Ala	Lys	Arg	Gln	Val	Ala	Leu	Trp	Thr	Leu	Gly	Gln	Leu	
				835					840					845			
30	GTG	GCC	AGC	ACT	GGC	TAT	GTA	GTA	GAG	CCC	TAC	AGG	AAG	TAC	CCT	ACT	2592
	Val	Ala	Ser	Thr	Gly	Tyr	Val	Val	Glu	Pro	Tyr	Arg	Lys	Tyr	Pro	Thr	
			850					855					860				
35	TTG	CTT	GAG	GTG	CTA	CTG	AAT	TTT	CTG	AAG	ACT	GAG	CAG	AAC	CAG	GGT	2640
	Leu	Leu	Glu	Val	Leu	Leu	Asn	Phe	Leu	Lys	Thr	Glu	Gln	Asn	Gln	Gly	
		865					870					875					
40	ACA	CGC	AGA	GAG	GCC	ATC	CGT	GTG	TTA	GGG	CTT	TTA	GGG	GCT	TTG	GAT	2688
	Thr	Arg	Arg	Glu	Ala	Ile	Arg	Val	Leu	Gly	Leu	Leu	Gly	Ala	Leu	Asp	
	880					885				890						895	
45	CCT	TAC	AAG	CAC	AAA	GTG	AAC	ATT	GGC	ATG	ATA	GAC	CAG	TCC	CGG	GAT	2736
	Pro	Tyr	Lys	His	Lys	Val	Asn	Ile	Gly	MET	Ile	Asp	Gln	Ser	Arg	Asp	
					900					905					910		
50	GCC	TCT	GCT	GTC	AGC	CTG	TCA	GAA	TCC	AAG	TCA	AGT	CAG	GAT	TCC	TCT	2784
	Ala	Ser	Ala	Val	Ser	Leu	Ser	Glu	Ser	Lys	Ser	Ser	Gln	Asp	Ser	Ser	
				915					920					925			
55	GAC	TAT	AGC	ACT	AGT	GAA	ATG	CTG	GTC	AAC	ATG	GGA	AAC	TTG	CCT	CTG	2832
	Asp	Tyr	Ser	Thr	Ser	Glu	MET	Leu	Val	Asn	MET	Gly	Asn	Leu	Pro	Leu	
			930					935					940				
60	GAT	GAG	TTC	TAC	CCA	GCT	GTG	TCC	ATG	GTG	GCC	CTG	ATG	CGG	ATC	TTC	2880
	Asp	Glu	Phe	Tyr	Pro	Ala	Val	Ser	MET	Val	Ala	Leu	MET	Arg	Ile	Phe	
		945					950					955					

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	CGA	GAC	CAG	TCA	CTC	TCT	CAT	CAT	CAC	ACC	ATG	GTT	GTC	CAG	GCC	ATC	2928
	Arg	Asp	Gln	Ser	Leu	Ser	His	His	His	Thr	MET	Val	Val	Gln	Ala	Ile	
	960					965					970					975	
5	ACC	TTC	ATC	TTC	AAG	TCC	CTG	GGA	CTC	AAA	TGT	GTG	CAG	TTC	CTG	CCC	2976
	Thr	Phe	Ile	Phe	Lys	Ser	Leu	Gly	Leu	Lys	Cys	Val	Gln	Phe	Leu	Pro	
					980					985						990	
10	CAG	GTC	ATG	CCC	ACG	TTC	CTT	AAT	GTC	ATT	CGA	GTC	TGT	GAT	GGG	GCC	3024
	Gln	Val	MET	Pro	Thr	Phe	Leu	Asn	Val	Ile	Arg	Val	Cys	Asp	Gly	Ala	
				995					1000						1005		
15	ATC	CGG	GAA	TTT	TTG	TTC	CAG	CAG	CTG	GGA	ATG	TTG	GTG	TCC	TTT	GTG	3072
	Ile	Arg	Glu	Phe	Leu	Phe	Gln	Gln	Leu	Gly	MET	Leu	Val	Ser	Phe	Val	
			1010						1015					1020			
20	AAG	AGC	CAC	ATC	AGA	CCT	TAT	ATG	GAT	GAA	ATA	GTC	ACC	CTC	ATG	AGA	3120
	Lys	Ser	His	Ile	Arg	Pro	Tyr	MET	Asp	Glu	Ile	Val	Thr	Leu	MET	Arg	
		1025					1030					1035					
25	GAA	TTC	TGG	GTC	ATG	AAC	ACC	TCA	ATT	CAG	AGC	ACG	ATC	ATT	CTT	CTC	3168
	Glu	Phe	Trp	Val	MET	Asn	Thr	Ser	Ile	Gln	Ser	Thr	Ile	Ile	Leu	Leu	
	1040					1045					1050					1055	
30	ATT	GAG	CAA	ATT	GTG	GTA	GCT	CTT	GGG	GGT	GAA	TTT	AAG	CTC	TAC	CTG	3216
	Ile	Glu	Gln	Ile	Val	Val	Ala	Leu	Gly	Gly	Glu	Phe	Lys	Leu	Tyr	Leu	
					1060					1065					1070		
35	CCC	CAG	CTG	ATC	CCA	CAC	ATG	CTG	CGT	GTC	TTC	ATG	CAT	GAC	AAC	AGC	3264
	Pro	Gln	Leu	Ile	Pro	His	MET	Leu	Arg	Val	Phe	MET	His	Asp	Asn	Ser	
				1075					1080						1085		
40	CCA	GGC	CGC	ATT	GTC	TCT	ATC	AAG	TTA	CTG	GCT	GCA	ATC	CAG	CTG	TTT	3312
	Pro	Gly	Arg	Ile	Val	Ser	Ile	Lys	Leu	Leu	Ala	Ala	Ile	Gln	Leu	Phe	
			1090					1095						1100			
45	GGC	GCC	AAC	CTG	GAT	GAC	TAC	CTG	CAT	TTA	CTG	CTG	CCT	CCT	ATT	GTT	3360
	Gly	Ala	Asn	Leu	Asp	Asp	Tyr	Leu	His	Leu	Leu	Leu	Pro	Pro	Ile	Val	
		1105					1110						1115				
50	AAG	TTG	TTT	GAT	GCC	CCT	GAA	GCT	CCA	CTG	CCA	TCT	CGA	AAG	GCA	GCG	3408
	Lys	Leu	Phe	Asp	Ala	Pro	Glu	Ala	Pro	Leu	Pro	Ser	Arg	Lys	Ala	Ala	
	1120					1125					1130					1135	
55	CTA	GAG	ACT	GTG	GAC	CGC	CTG	ACG	GAG	TCC	CTG	GAT	TTC	ACT	GAC	TAT	3456
	Leu	Glu	Thr	Val	Asp	Arg	Leu	Thr	Glu	Ser	Leu	Asp	Phe	Thr	Asp	Tyr	
					1140					1145					1150		
60	GCC	TCC	CGG	ATC	ATT	CAC	CCT	ATT	GTT	CGA	ACA	CTG	GAC	CAG	AGC	CCA	3504
	Ala	Ser	Arg	Ile	Ile	His	Pro	Ile	Val	Arg	Thr	Leu	Asp	Gln	Ser	Pro	
				1155					1160					1165			

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	GAA CTG CGC TCC ACA GCC ATG GAC ACG CTG TCT TCA CTT GTT TTT CAG	3552
	Glu Leu Arg Ser Thr Ala MET Asp Thr Leu Ser Ser Leu Val Phe Gln	
	1170 1175 1180	
5	CTG GGG AAG AAG TAC CAA ATT TTC ATT CCA ATG GTG AAT AAA GTT CTG	3600
	Leu Gly Lys Lys Tyr Gln Ile Phe Ile Pro MET Val Asn Lys Val Leu	
	1185 1190 1195	
10	GTG CGA CAC CGA ATC AAT CAT CAG CGC TAT GAT GTG CTC ATC TGC AGA	3648
	Val Arg His Arg Ile Asn His Gln Arg Tyr Asp Val Leu Ile Cys Arg	
	1200 1205 1210 1215	
15	ATT GTC AAG GGA TAC ACA CTT GCT GAT GAA GAG GAG GAT CCT TTG ATT	3696
	Ile Val Lys Gly Tyr Thr Leu Ala Asp Glu Glu Glu Asp Pro Leu Ile	
	1220 1225 1230	
20	TAC CAG CAT CGG ATG CTT AGG AGT GGC CAA GGG GAT GCA TTG GCT AGT	3744
	Tyr Gln His Arg MET Leu Arg Ser Gly Gln Gly Asp Ala Leu Ala Ser	
	1235 1240 1245	
25	GGA CCA GTG GAA ACA GGA CCC ATG AAG AAA CTG CAC GTC AGC ACC ATC	3792
	Gly Pro Val Glu Thr Gly Pro MET Lys Lys Leu His Val Ser Thr Ile	
	1250 1255 1260	
30	AAC CTC CAA AAG GCC TGG GGC GCT GCC AGG AGG GTC TCC AAA GAT GAC	3840
	Asn Leu Gln Lys Ala Trp Gly Ala Ala Arg Arg Val Ser Lys Asp Asp	
	1265 1270 1275	
35	TGG CTG GAA TGG CTG AGA CGG CTG AGC CTG GAG CTG CTG AAG GAC TCA	3888
	Trp Leu Glu Trp Leu Arg Arg Leu Ser Leu Glu Leu Leu Lys Asp Ser	
	1280 1285 1290 1295	
40	TCA TCG CCC TCC CTG CGC TCC TGC TGG GCC CTG GCA CAG GCC TAC AAC	3936
	Ser Ser Pro Ser Leu Arg Ser Cys Trp Ala Leu Ala Gln Ala Tyr Asn	
	1300 1305 1310	
45	CCG ATG GCC AGG GAT CTC TTC AAT GCT GCA TTT GTG TCC TGC TGG TCT	3984
	Pro MET Ala Arg Asp Leu Phe Asn Ala Ala Phe Val Ser Cys Trp Ser	
	1315 1320 1325	
50	GAA CTG AAT GAA GAT CAA CAG GAT GAG CTC ATC AGA AGC ATC GAG TTG	4032
	Glu Leu Asn Glu Asp Gln Gln Asp Glu Leu Ile Arg Ser Ile Glu Leu	
	1330 1335 1340	
55	GCC CTC ACC TCA CAA GAC ATC GCT GAA GTC ACA CAG ACC CTC TTA AAC	4080
	Ala Leu Thr Ser Gln Asp Ile Ala Glu Val Thr Gln Thr Leu Leu Asn	
	1345 1350 1355	
60	TTG GCT GAA TTC ATG GAA CAC AGT GAC AAG GGC CCC CTG CCA CTG AGA	4128
	Leu Ala Glu Phe MET Glu His Ser Asp Lys Gly Pro Leu Pro Leu Arg	
	1360 1365 1370 1375	

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	GAT GAC AAT GGC ATT GTT CTG CTG GGT GAG AGA GCT GCC AAG TGC CGA	4176
	Asp Asp Asn Gly Ile Val Leu Leu Gly Glu Arg Ala Ala Lys Cys Arg	
	1380 1385 1390	
5	GCA TAT GCC AAA GCA CTA CAC TAC AAA GAA CTG GAG TTC CAG AAA GGC	4224
	Ala Tyr Ala Lys Ala Leu His Tyr Lys Glu Leu Glu Phe Gln Lys Gly	
	1395 1400 1405	
10	CCC ACC CCT GCC ATT CTA GAA TCT CTC ATC AGC ATT AAT AAT AAG CTA	4272
	Pro Thr Pro Ala Ile Leu Glu Ser Leu Ile Ser Ile Asn Asn Lys Leu	
	1410 1415 1420	
15	CAG CAG CCG GAG GCA GCG GCC GGA GTG TTA GAA TAT GCC ATG AAA CAC	4320
	Gln Gln Pro Glu Ala Ala Ala Gly Val Leu Glu Tyr Ala MET Lys His	
	1425 1430 1435	
20	TTT GGA GAG CTG GAG ATC CAG GCT ACC TGG TAT GAG AAA CTG CAC GAG	4368
	Phe Gly Glu Leu Glu Ile Gln Ala Thr Trp Tyr Glu Lys Leu His Glu	
	1440 1445 1450 1455	
	TGG GAG GAT GCC CTT GTG GCC TAT GAC AAG AAA ATG GAC ACC AAC AAG	4416
	Trp Glu Asp Ala Leu Val Ala Tyr Asp Lys Lys MET Asp Thr Asn Lys	
	1460 1465 1470	
25	GAC GAC CCA GAG CTG ATG CTG GGC CGC ATG CGC TGC CTC GAG GCC TTG	4464
	Asp Asp Pro Glu Leu MET Leu Gly Arg MET Arg Cys Leu Glu Ala Leu	
	1475 1480 1485	
30	GGG GAA TGG GGT CAA CTC CAC CAG CAG TGC TGT GAA AAG TGG ACC CTG	4512
	Gly Glu Trp Gly Gln Leu His Gln Gln Cys Cys Glu Lys Trp Thr Leu	
	1490 1495 1500	
35	GTT AAT GAT GAG ACC CAA GCC AAG ATG GCC CGG ATG GCT GCT GCA GCT	4560
	Val Asn Asp Glu Thr Gln Ala Lys MET Ala Arg MET Ala Ala Ala Ala	
	1505 1510 1515	
40	GCA TGG GGT TTA GGT CAG TGG GAC AGC ATG GAA GAA TAC ACC TGT ATG	4608
	Ala Trp Gly Leu Gly Gln Trp Asp Ser MET Glu Glu Tyr Thr Cys MET	
	1520 1525 1530 1535	
	ATC CCT CGG GAC ACC CAT GAT GGG GCA TTT TAT AGA GCT GTG CTG GCA	4656
	Ile Pro Arg Asp Thr His Asp Gly Ala Phe Tyr Arg Ala Val Leu Ala	
	1540 1545 1550	
45	CTG CAT CAG GAC CTC TTC TCC TTG GCA CAA CAG TGC ATT GAC AAG GCC	4704
	Leu His Gln Asp Leu Phe Ser Leu Ala Gln Gln Cys Ile Asp Lys Ala	
	1555 1560 1565	
50	AGG GAC CTG CTG GAT GCT GAA TTA ACT GCA ATG GCA GGA GAG AGT TAC	4752
	Arg Asp Leu Leu Asp Ala Glu Leu Thr Ala MET Ala Gly Glu Ser Tyr	
	1570 1575 1580	

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	AGT	CGG	GCA	TAT	GGG	GCC	ATG	GTT	TCT	TGC	CAC	ATG	CTG	TCC	GAG	CTG	4800
	Ser	Arg	Ala	Tyr	Gly	Ala	MET	Val	Ser	Cys	His	MET	Leu	Ser	Glu	Leu	
		1585					1590					1595					
5	GAG	GAG	GTT	ATC	CAG	TAC	AAA	CTT	GTC	CCC	GAG	CGA	CGA	GAG	ATC	ATC	4848
	Glu	Glu	Val	Ile	Gln	Tyr	Lys	Leu	Val	Pro	Glu	Arg	Arg	Glu	Ile	Ile	
	1600				1605						1610				1615		
10	CGC	CAG	ATC	TGG	TGG	GAG	AGA	CTG	CAG	GGC	TGC	CAG	CGT	ATC	GTA	GAG	4896
	Arg	Gln	Ile	Trp	Trp	Glu	Arg	Leu	Gln	Gly	Cys	Gln	Arg	Ile	Val	Glu	
				1620						1625					1630		
15	GAC	TGG	CAG	AAA	ATC	CTT	ATG	GTG	CGG	TCC	CTT	GTG	GTC	AGC	CCT	CAT	4944
	Asp	Trp	Gln	Lys	Ile	Leu	MET	Val	Arg	Ser	Leu	Val	Val	Ser	Pro	His	
			1635						1640					1645			
20	GAA	GAC	ATG	AGA	ACC	TGG	CTC	AAG	TAT	GCA	AGC	CTG	TGC	GGC	AAG	AGT	4992
	Glu	Asp	MET	Arg	Thr	Trp	Leu	Lys	Tyr	Ala	Ser	Leu	Cys	Gly	Lys	Ser	
			1650					1655					1660				
25	GGC	AGG	CTG	GCT	CTT	GCT	CAT	AAA	ACT	TTA	GTG	TTG	CTC	CTG	GGA	GTT	5040
	Gly	Arg	Leu	Ala	Leu	Ala	His	Lys	Thr	Leu	Val	Leu	Leu	Leu	Gly	Val	
		1665				1670					1675						
30	GAT	CCG	TCT	CGG	CAA	CTT	GAC	CAT	CCT	CTG	CCA	ACA	GTT	CAC	CCT	CAG	5088
	Asp	Pro	Ser	Arg	Gln	Leu	Asp	His	Pro	Leu	Pro	Thr	Val	His	Pro	Gln	
		1680			1685						1690					1695	
35	GTG	ACC	TAT	GCC	TAC	ATG	AAA	AAC	ATG	TGG	AAG	AGT	GCC	CGC	AAG	ATC	5136
	Val	Thr	Tyr	Ala	Tyr	MET	Lys	Asn	MET	Trp	Lys	Ser	Ala	Arg	Lys	Ile	
				1700						1705					1710		
40	GAT	GCC	TTC	CAG	CAC	ATG	CAG	CAT	TTT	GTC	CAG	ACC	ATG	CAG	CAA	CAG	5184
	Asp	Ala	Phe	Gln	His	MET	Gln	His	Phe	Val	Gln	Thr	MET	Gln	Gln	Gln	
			1715						1720					1725			
45	GCC	CAG	CAT	GCC	ATC	GCT	ACT	GAG	GAC	CAG	CAG	CAT	AAG	CAG	GAA	CTG	5232
	Ala	Gln	His	Ala	Ile	Ala	Thr	Glu	Asp	Gln	Gln	His	Lys	Gln	Glu	Leu	
			1730					1735					1740				
50	CAC	AAG	CTC	ATG	GCC	CGA	TGC	TTC	CTG	AAA	CTT	GGA	GAG	TGG	CAG	CTG	5280
	His	Lys	Leu	MET	Ala	Arg	Cys	Phe	Leu	Lys	Leu	Gly	Glu	Trp	Gln	Leu	
		1745				1750						1755					
55	AAT	CTA	CAG	GGC	ATC	AAT	GAG	AGC	ACA	ATC	CCC	AAA	GTG	CTG	CAG	TAC	5328
	Asn	Leu	Gln	Gly	Ile	Asn	Glu	Ser	Thr	Ile	Pro	Lys	Val	Leu	Gln	Tyr	
	1760				1765						1770				1775		
60	TAC	AGC	GCC	GCC	ACA	GAG	CAC	GAC	CGC	AGC	TGG	TAC	AAG	GCC	TGG	CAT	5376
	Tyr	Ser	Ala	Ala	Thr	Glu	His	Asp	Arg	Ser	Trp	Tyr	Lys	Ala	Trp	His	
					1780					1785					1790		

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	GCG	TGG	GCA	GTG	ATG	AAC	TTC	GAA	GCT	GTG	CTA	CAC	TAC	AAA	CAT	CAG	5424
	Ala	Trp	Ala	Val	MET	Asn	Phe	Glu	Ala	Val	Leu	His	Tyr	Lys	His	Gln	
				1795					1800					1805			
5	AAC	CAA	GCC	CGC	GAT	GAG	AAG	AAG	AAA	CTG	CGT	CAT	GCC	AGC	GGG	GCC	5472
	Asn	Gln	Ala	Arg	Asp	Glu	Lys	Lys	Lys	Leu	Arg	His	Ala	Ser	Gly	Ala	
			1810					1815					1820				
10	AAC	ATC	ACC	AAC	GCC	ACC	ACT	GCC	GCC	ACC	ACG	GCC	GCC	ACT	GCC	ACC	5520
	Asn	Ile	Thr	Asn	Ala	Thr	Thr	Ala	Ala	Thr	Thr	Ala	Ala	Thr	Ala	Thr	
			1825					1830				1835					
15	ACC	ACT	GCC	AGC	ACC	GAG	GGC	AGC	AAC	AGT	GAG	AGC	GAG	GCC	GAG	AGC	5568
	Thr	Thr	Ala	Ser	Thr	Glu	Gly	Ser	Asn	Ser	Glu	Ser	Glu	Ala	Glu	Ser	
	1840					1845				1850					1855		
20	ACC	GAG	AAC	AGC	CCC	ACC	CCA	TCG	CCG	CTG	CAG	AAG	AAG	GTC	ACT	GAG	5616
	Thr	Glu	Asn	Ser	Pro	Thr	Pro	Ser	Pro	Leu	Gln	Lys	Lys	Val	Thr	Glu	
					1860					1865					1870		
25	GAT	CTG	TCC	AAA	ACC	CTC	CTG	ATG	TAC	ACG	GTG	CCT	GCC	GTC	CAG	GGC	5664
	Asp	Leu	Ser	Lys	Thr	Leu	Leu	MET	Tyr	Thr	Val	Pro	Ala	Val	Gln	Gly	
				1875					1880					1885			
30	TTC	TTC	CGT	TCC	ATC	TCC	TTG	TCA	CGA	GGC	AAC	AAC	CTC	CAG	GAT	ACA	5712
	Phe	Phe	Arg	Ser	Ile	Ser	Leu	Ser	Arg	Gly	Asn	Asn	Leu	Gln	Asp	Thr	
			1890					1895					1900				
35	CTC	AGA	GTT	CTC	ACC	TTA	TGG	TTT	GAT	TAT	GGT	CAC	TGG	CCA	GAT	GTC	5760
	Leu	Arg	Val	Leu	Thr	Leu	Trp	Phe	Asp	Tyr	Gly	His	Trp	Pro	Asp	Val	
		1905					1910					1915					
40	AAT	GAG	GCC	TTA	GTG	GAG	GGG	GTG	AAA	GCC	ATC	CAG	ATT	GAT	ACC	TGG	5808
	Asn	Glu	Ala	Leu	Val	Glu	Gly	Val	Lys	Ala	Ile	Gln	Ile	Asp	Thr	Trp	
	1920					1925				1930						1935	
45	CTA	CAG	GTT	ATA	CCT	CAG	CTC	ATT	GCA	AGA	ATT	GAT	ACG	CCC	AGA	CCC	5856
	Leu	Gln	Val	Ile	Pro	Gln	Leu	Ile	Ala	Arg	Ile	Asp	Thr	Pro	Arg	Pro	
					1940					1945					1950		
50	TTG	GTG	GGA	CGT	CTC	ATT	CAC	CAG	CTT	CTC	ACA	GAC	ATT	GGT	CGG	TAC	5904
	Leu	Val	Gly	Arg	Leu	Ile	His	Gln	Leu	Leu	Thr	Asp	Ile	Gly	Arg	Tyr	
				1955				1960						1965			
55	CAC	CCC	CAG	GCC	CTC	ATC	TAC	CCA	CTG	ACA	GTG	GCT	TCT	AAG	TCT	ACC	5952
	His	Pro	Gln	Ala	Leu	Ile	Tyr	Pro	Leu	Thr	Val	Ala	Ser	Lys	Ser	Thr	
			1970					1975					1980				
60	ACG	ACA	GCC	CGG	CAC	AAT	GCA	GCC	AAC	AAG	ATT	CTG	AAG	AAC	ATG	TGT	6000
	Thr	Thr	Ala	Arg	His	Asn	Ala	Ala	Asn	Lys	Ile	Leu	Lys	Asn	MET	Cys	
			1985				1990					1995					

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	GAG CAC AGC AAC ACC CTG GTC CAG CAG GCC ATG ATG GTG AGC GAG GAG	6048
	Glu His Ser Asn Thr Leu Val Gln Gln Ala MET MET Val Ser Glu Glu	
	2000 2005 2010 2015	
5	CTG ATC CGA GTG GCC ATC CTC TGG CAT GAG ATG TGG CAT GAA GGC CTG	6096
	Leu Ile Arg Val Ala Ile Leu Trp His Glu MET Trp His Glu Gly Leu	
	2020 2025 2030	
10	GAA GAG GCA TCT CGT TTG TAC TTT GGG GAA AGG AAC GTG AAA GGC ATG	6144
	Glu Glu Ala Ser Arg Leu Tyr Phe Gly Glu Arg Asn Val Lys Gly MET	
	2035 2040 2045	
15	TTT GAG GTG CTG GAG CCC TTG CAT GCT ATG ATG GAA CGG GGC CCC CAG	6192
	Phe Glu Val Leu Glu Pro Leu His Ala MET MET Glu Arg Gly Pro Gln	
	2050 2055 2060	
	ACT CTG AAG GAA ACA TCC TTT AAT CAG GCC TAT GGT CGA GAT TTA ATG	6240
	Thr Leu Lys Glu Thr Ser Phe Asn Gln Ala Tyr Gly Arg Asp Leu MET	
	2065 2070 2075	
20	GAG GCC CAA GAG TGG TGC AGG AAG TAC ATG AAA TCA GGG AAT GTC AAG	6288
	Glu Ala Gln Glu Trp Cys Arg Lys Tyr MET Lys Ser Gly Asn Val Lys	
	2080 2085 2090 2095	
25	GAC CTC ACC CAA GCC TGG GAC CTC TAT TAT CAT GTG TTC CGA CGA ATC	6336
	Asp Leu Thr Gln Ala Trp Asp Leu Tyr Tyr His Val Phe Arg Arg Ile	
	2100 2105 2110	
30	TCA AAG CAG CTG CCT CAG CTC ACA TCC TTA GAG CTG CAA TAT GTT TCC	6384
	Ser Lys Gln Leu Pro Gln Leu Thr Ser Leu Glu Leu Gln Tyr Val Ser	
	2115 2120 2125	
35	CCA AAA CTT CTG ATG TGC CGG GAC CTT GAA TTG GCT GTG CCA GGA ACA	6432
	Pro Lys Leu Leu MET Cys Arg Asp Leu Glu Leu Ala Val Pro Gly Thr	
	2130 2135 2140	
	TAT GAC CCC AAC CAG CCA ATC ATT CGC ATT CAG TCC ATA GCA CCG TCT	6480
	Tyr Asp Pro Asn Gln Pro Ile Ile Arg Ile Gln Ser Ile Ala Pro Ser	
	2145 2150 2155	
40	TTG CAA GTC ATC ACA TCC AAG CAG AGG CCC CGG AAA TTG ACA CTT ATG	6528
	Leu Gln Val Ile Thr Ser Lys Gln Arg Pro Arg Lys Leu Thr Leu MET	
	2160 2165 2170 2175	
45	GGC AGC AAC GGA CAT GAG TTT GTT TTC CTT CTA AAA GGC CAT GAA GAT	6576
	Gly Ser Asn Gly His Glu Phe Val Phe Leu Leu Lys Gly His Glu Asp	
	2180 2185 2190	
50	CTG CGC CAG GAT GAG CGT GTG ATG CAG CTC TTC GGC CTG GTT AAC ACC	6624
	Leu Arg Gln Asp Glu Arg Val MET Gln Leu Phe Gly Leu Val Asn Thr	
	2195 2200 2205	

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	CTT	CTG	GCC	AAT	GAC	CCA	ACA	TCT	CTT	CGG	AAA	AAC	CTC	AGC	ATC	CAG	6672
	Leu	Leu	Ala	Asn	Asp	Pro	Thr	Ser	Leu	Arg	Lys	Asn	Leu	Ser	Ile	Gln	
			2210					2215					2220				
5	AGA	TAC	GCT	GTC	ATC	CCT	TTA	TCG	ACC	AAC	TCG	GGC	CTC	ATT	GGC	TGG	6720
	Arg	Tyr	Ala	Val	Ile	Pro	Leu	Ser	Thr	Asn	Ser	Gly	Leu	Ile	Gly	Trp	
		2225					2230					2235					
10	GTT	CCC	CAC	TGT	GAC	ACA	CTG	CAC	GCC	CTC	ATC	CGG	GAC	TAC	AGG	GAG	6768
	Val	Pro	His	Cys	Asp	Thr	Leu	His	Ala	Leu	Ile	Arg	Asp	Tyr	Arg	Glu	
	2240					2245					2250					2255	
15	AAG	AAG	AAG	ATC	CTT	CTC	AAC	ATC	GAG	CAT	CGC	ATC	ATG	TTG	CGG	ATG	6816
	Lys	Lys	Lys	Ile	Leu	Leu	Asn	Ile	Glu	His	Arg	Ile	MET	Leu	Arg	MET	
				2260					2265						2270		
20	GCT	CCG	GAC	TAT	GAC	CAC	TTG	ACT	CTG	ATG	CAG	AAG	GTG	GAG	GTG	TTT	6864
	Ala	Pro	Asp	Tyr	Asp	His	Leu	Thr	Leu	MET	Gln	Lys	Val	Glu	Val	Phe	
				2275					2280					2285			
25	GAG	CAT	GCC	GTC	AAT	AAT	ACA	GCT	GGG	GAC	GAC	CTG	GCC	AAG	CTG	CTG	6912
	Glu	His	Ala	Val	Asn	Asn	Thr	Ala	Gly	Asp	Asp	Leu	Ala	Lys	Leu	Leu	
			2290				2295					2300					
30	TGG	CTG	AAA	AGC	CCC	AGC	TCC	GAG	GTG	TGG	TTT	GAC	CGA	AGA	ACC	AAT	6960
	Trp	Leu	Lys	Ser	Pro	Ser	Ser	Glu	Val	Trp	Phe	Asp	Arg	Arg	Thr	Asn	
		2305					2310					2315					
35	TAT	ACC	CGT	TCT	TTA	GCG	GTC	ATG	TCA	ATG	GTT	GGG	TAT	ATT	TTA	GGC	7008
	Tyr	Thr	Arg	Ser	Leu	Ala	Val	MET	Ser	MET	Val	Gly	Tyr	Ile	Leu	Gly	
	2320					2325					2330					2335	
40	CTG	GGA	GAT	AGA	CAC	CCA	TCC	AAC	CTG	ATG	CTG	GAC	CGT	CTG	AGT	GGG	7056
	Leu	Gly	Asp	Arg	His	Pro	Ser	Asn	Leu	MET	Leu	Asp	Arg	Leu	Ser	Gly	
				2340					2345						2350		
45	AAG	ATC	CTG	CAC	ATT	GAC	TTT	GGG	GAC	TGC	TTT	GAG	GTT	GCT	ATG	ACC	7104
	Lys	Ile	Leu	His	Ile	Asp	Phe	Gly	Asp	Cys	Phe	Glu	Val	Ala	MET	Thr	
				2355				2360						2365			
50	CGA	GAG	AAG	TTT	CCA	GAG	AAG	ATT	CCA	TTT	AGA	CTA	ACA	AGA	ATG	TTG	7152
	Arg	Glu	Lys	Phe	Pro	Glu	Lys	Ile	Pro	Phe	Arg	Leu	Thr	Arg	MET	Leu	
			2370				2375					2380					
55	ACC	AAT	GCT	ATG	GAG	GTT	ACA	GGC	CTG	GAT	GGC	AAC	TAC	AGA	ATC	ACA	7200
	Thr	Asn	Ala	MET	Glu	Val	Thr	Gly	Leu	Asp	Gly	Asn	Tyr	Arg	Ile	Thr	
		2385					2390					2395					
60	TGC	CAC	ACA	GTG	ATG	GAG	GTG	CTG	CGA	GAG	CAC	AAG	GAC	AGT	GTC	ATG	7248
	Cys	His	Thr	Val	MET	Glu	Val	Leu	Arg	Glu	His	Lys	Asp	Ser	Val	MET	
	2400					2405					2410					2415	

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GCC GTG CTG GAA GCC TTT GTC TAT GAC CCC TTG CTG AAC TGG AGG CTG ' 7296
Ala Val Leu Glu Ala Phe Val Tyr Asp Pro Leu Leu Asn Trp Arg Leu
                                2420                                2425                                2430

5  ATG GAC ACA AAT ACC AAA GGC AAC AAG CGA TCC CGA ACG AGG ACG GAT 7344
MET Asp Thr Asn Thr Lys Gly Asn Lys Arg Ser Arg Thr Arg Thr Asp
                                2435                                2440                                2445

10 TCC TAC TCT GCT GGC CAG TCA GTC GAA ATT TTG GAC GGT GTG GAA CTT 7392
Ser Tyr Ser Ala Gly Gln Ser Val Glu Ile Leu Asp Gly Val Glu Leu
                                2450                                2455                                2460

GGA GAG CCA GCC CAT AAG AAA ACG GGG ACC ACA GTG CCA GAA TCT ATT ' 7440
Gly Glu Pro Ala His Lys Lys Thr Gly Thr Thr Val Pro Glu Ser Ile
15 2465                                2470                                2475

CAT TCT TTC ATT GGA GAC GGT TTG GTG AAA CCA GAG GCC CTA AAT AAG 7488
His Ser Phe Ile Gly Asp Gly Leu Val Lys Pro Glu Ala Leu Asn Lys
2480                                2485                                2490                                2495

20 AAA GCT ATC CAG ATT ATT AAC AGG GTT CGA GAT AAG CTC ACT GGT CGG 7536
Lys Ala Ile Gln Ile Ile Asn Arg Val Arg Asp Lys Leu Thr Gly Arg
                                2500                                2505                                2510

25 GAC TTC TCT CAT GAT GAC ACT TTG GAT GTT CCA ACG CAA GTT GAG CTG 7584
Asp Phe Ser His Asp Asp Thr Leu Asp Val Pro Thr Gln Val Glu Leu
                                2515                                2520                                2525

30 CTC ATC AAA CAA GCG ACA TCC CAT GAA AAC CTC TGC CAG TGC TAT ATT 7632
Leu Ile Lys Gln Ala Thr Ser His Glu Asn Leu Cys Gln Cys Tyr Ile
                                2530                                2535                                2540

GCG TGG TAC CCT TTC TGG TAA 7653
Gly Trp Tyr Pro Phe Trp
35 2545

(3) INFORMATION FOR SEQ. ID NO: 2:
    (i) SEQUENCE CHARACTERISTICS:
        (A) LENGTH: 3423
        (B) TYPE: nucleic acid
        (C) STRANDEDNESS: double-stranded
        (D) TOPOLOGY: linear
    40

    (ii) MOLECULE TYPE: sense orientation of double-stranded
        cDNA to mRNA
        (A) DESCRIPTION: Sequence No. 2 illustrates a
        45 GST-SEP45 fusion protein beginning
        at the first amino acid of the GST-SEP45
        protein.
        50

    (iii) HYPOTHETICAL: no
    (iv) ANTISENSE: no
    (vi) ORIGINAL SOURCE:
        (A) ORGANISM: Molt 4 human T-cell leukemia cells
        55 (B) STRAIN: ATCC Strain CRL 1582
  
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(xi)		SEQUENCE DESCRIPTION: SEQ. ID NO: 2															
5	ATG	TCC	CCT	ATA	CTA	GGT	TAT	TGG	AAA	ATT	AAG	GGC	CTT	GTG	CAA	CCC	48
	MET	Ser	Pro	Ile	Leu	Gly	Tyr	Trp	Lys	Ile	Lys	Gly	Leu	Val	Gln	Pro	
	1				5					10					15		
10	ACT	CGA	CTT	CTT	TTG	GAA	TAT	CTT	GAA	GAA	AAA	TAT	GAA	GAG	CAT	TTG	96
	Thr	Arg	Leu	Leu	Leu	Glu	Tyr	Leu	Glu	Glu	Lys	Tyr	Glu	Glu	His	Leu	
				20					25					30			
15	TAT	GAG	CGC	GAT	GAA	GGT	GAT	AAA	TGG	CGA	AAC	AAA	AAG	TTT	GAA	TTG	144
	Tyr	Glu	Arg	Asp	Glu	Gly	Asp	Lys	Trp	Arg	Asn	Lys	Lys	Phe	Glu	Leu	
			35					40					45				
20	GGT	TTG	GAG	TTT	CCC	AAT	CTT	CCT	TAT	TAT	ATT	GAT	GGT	GAT	GTT	AAA	192
	Gly	Leu	Glu	Phe	Pro	Asn	Leu	Pro	Tyr	Tyr	Ile	Asp	Gly	Asp	Val	Lys	
			50				55					60					
25	TTA	ACA	CAG	TCT	ATG	GCC	ATC	ATA	CGT	TAT	ATA	GCT	GAC	AAG	CAC	AAC	240
	Leu	Thr	Gln	Ser	MET	Ala	Ile	Ile	Arg	Tyr	Ile	Ala	Asp	Lys	His	Asn	
	65					70					75					80	
30	ATG	TTG	GGT	GGT	TGT	CCA	AAA	GAG	CGT	GCA	GAG	ATT	TCA	ATG	CTT	GAA	288
	MET	Leu	Gly	Gly	Cys	Pro	Lys	Glu	Arg	Ala	Glu	Ile	Ser	MET	Leu	Glu	
					85					90					95		
35	GGA	GCG	GTT	TTG	GAT	ATT	AGA	TAC	GGT	GTT	TCG	AGA	ATT	GCA	TAT	AGT	336
	Gly	Ala	Val	Leu	Asp	Ile	Arg	Tyr	Gly	Val	Ser	Arg	Ile	Ala	Tyr	Ser	
				100					105					110			
40	AAA	GAC	TTT	GAA	ACT	CTC	AAA	GTT	GAT	TTT	CTT	AGC	AAG	CTA	CCT	GAA	384
	Lys	Asp	Phe	Glu	Thr	Leu	Lys	Val	Asp	Phe	Leu	Ser	Lys	Leu	Pro	Glu	
			115					120					125				
45	ATG	CTG	AAA	ATG	TTC	GAA	GAT	CGT	TTA	TGT	CAT	AAA	ACA	TAT	TTA	AAT	432
	MET	Leu	Lys	MET	Phe	Glu	Asp	Arg	Leu	Cys	His	Lys	Thr	Tyr	Leu	Asn	
			130				135					140					
50	GGT	GAT	CAT	GTA	ACC	CAT	CCT	GAC	TTC	ATG	TTG	TAT	GAC	GCT	CTT	GAT	480
	Gly	Asp	His	Val	Thr	His	Pro	Asp	Phe	MET	Leu	Tyr	Asp	Ala	Leu	Asp	
	145					150					155				160		
55	GTT	GTT	TTA	TAC	ATG	GAC	CCA	ATG	TGC	CTG	GAT	GCG	TTC	CCA	AAA	TTA	528
	Val	Val	Leu	Tyr	MET	Asp	Pro	MET	Cys	Leu	Asp	Ala	Phe	Pro	Lys	Leu	
					165					170					175		
60	GTT	TGT	TTT	AAA	AAA	CGT	ATT	GAA	GCT	ATC	CCA	CAA	ATT	GAT	AAG	TAC	576
	Val	Cys	Phe	Lys	Lys	Arg	Ile	Glu	Ala	Ile	Pro	Gln	Ile	Asp	Lys	Tyr	
				180					185					190			
65	TTG	AAA	TCC	AGC	AAG	TAT	ATA	GCA	TGG	CCT	TTG	CAG	GGC	TGG	CAA	GCC	624
	Leu	Lys	Ser	Ser	Lys	Tyr	Ile	Ala	Trp	Pro	Leu	Gln	Gly	Trp	Gln	Ala	
			195					200					205				

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	ACG	TTT	GGT	GGT	GGC	GAC	CAT	CCT	CCA	AAA	TCG	GAT	CTG	GTT	CCG	CGT	672
	Thr	Phe	Gly	Gly	Gly	Asp	His	Pro	Pro	Lys	Ser	Asp	Leu	Val	Pro	Arg	
	210						215					220					
5	GGT	GGA	TCC	CCG	GGA	ATT	TCC	GGT	GGT	GGT	GGT	GGT	GGA	ATT	CTA	GAC	720
	Gly	Gly	Ser	Pro	Gly	Ile	Ser	Gly	Gly	Gly	Gly	Gly	Gly	Ile	Leu	Asp	
	225					230					235					240	
10	GAC	TCC	ATG	AGC	TTC	AAG	TAT	GCA	AGC	CTG	TGC	GGC	AAG	AGT	GGC	AGG	768
	Asp	Ser	MET	Ser	Phe	Lys	Tyr	Ala	Ser	Leu	Cys	Gly	Lys	Ser	Gly	Arg	
					245					250					255		
15	CTG	GCT	CTT	GCT	CAT	AAA	ACT	TTA	GTG	TTG	CTC	CTG	GGA	GTT	GAT	CCG	816
	Leu	Ala	Leu	Ala	His	Lys	Thr	Leu	Val	Leu	Leu	Leu	Gly	Val	Asp	Pro	
				260					265					270			
20	TCT	CGG	CAA	CTT	GAC	CAT	CCT	CTG	CCA	ACA	GTT	CAC	CCT	CAG	GTG	ACC	864
	Ser	Arg	Gln	Leu	Asp	His	Pro	Leu	Pro	Thr	Val	His	Pro	Gln	Val	Thr	
			275					280					285				
25	TAT	GCC	TAC	ATG	AAA	AAC	ATG	TGG	AAG	AGT	GCC	CGC	AAG	ATC	GAT	GCC	912
	Tyr	Ala	Tyr	MET	Lys	Asn	MET	Trp	Lys	Ser	Ala	Arg	Lys	Ile	Asp	Ala	
		290					295				300						
30	TTC	CAG	CAC	ATG	CAG	CAT	TTT	GTC	CAG	ACC	ATG	CAG	CAA	CAG	GCC	CAG	960
	Phe	Gln	His	MET	Gln	His	Phe	Val	Gln	Thr	MET	Gln	Gln	Gln	Ala	Gln	
	305					310				315					320		
35	CAT	GCC	ATC	GCT	ACT	GAG	GAC	CAG	CAG	CAT	AAG	CAG	GAA	CTG	CAC	AAG	1008
	His	Ala	Ile	Ala	Thr	Glu	Asp	Gln	Gln	His	Lys	Gln	Glu	Leu	His	Lys	
					325					330					335		
40	CTC	ATG	GCC	CGA	TGC	TTC	CTG	AAA	CTT	GGA	GAG	TGG	CAG	CTG	AAT	CTA	1056
	Leu	MET	Ala	Arg	Cys	Phe	Leu	Lys	Leu	Gly	Glu	Trp	Gln	Leu	Asn	Leu	
				340				345						350			
45	CAG	GGC	ATC	AAT	GAG	AGC	ACA	ATC	CCC	AAA	GTG	CTG	CAG	TAC	TAC	AGC	1104
	Gln	Gly	Ile	Asn	Glu	Ser	Thr	Ile	Pro	Lys	Val	Leu	Gln	Tyr	Tyr	Ser	
			355					360					365				
50	GCC	GCC	ACA	GAG	CAC	GAC	CGC	AGC	TGG	TAC	AAG	GCC	TGG	CAT	GCG	TGG	1152
	Ala	Ala	Thr	Glu	His	Asp	Arg	Ser	Trp	Tyr	Lys	Ala	Trp	His	Ala	Trp	
		370					375					380					
55	GCA	GTG	ATG	AAC	TTC	GAA	GCT	GTG	CTA	CAC	TAC	AAA	CAT	CAG	AAC	CAA	1200
	Ala	Val	MET	Asn	Phe	Glu	Ala	Val	Leu	His	Tyr	Lys	His	Gln	Asn	Gln	
	385					390					395					400	
60	GCC	CGC	GAT	GAG	AAG	AAG	AAA	CTG	CGT	CAT	GCC	AGC	GGG	GCC	AAC	ATC	1248
	Ala	Arg	Asp	Glu	Lys	Lys	Lys	Leu	Arg	His	Ala	Ser	Gly	Ala	Asn	Ile	
					405				410						415		

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	ACC	AAC	GCC	ACC	ACT	GCC	GCC	ACC	ACG	GCC	GCC	ACT	GCC	ACC	ACC	ACT	1296
	Thr	Asn	Ala	Thr	Thr	Ala	Ala	Thr	Thr	Ala	Ala	Thr	Ala	Thr	Thr	Thr	
				420					425						430		
5	GCC	AGC	ACC	GAG	GGC	AGC	AAC	AGT	GAG	AGT	GAG	GCC	GAG	AGC	ACC	GAG	1344
	Ala	Ser	Thr	Glu	Gly	Ser	Asn	Ser	Glu	Ser	Glu	Ala	Glu	Ser	Thr	Glu	
			435					440					445				
10	AAC	AGC	CCC	ACC	CCA	TCG	CCG	CTG	CAG	AAG	AAG	GTC	ACT	GAG	GAT	CTG	1392
	Asn	Ser	Pro	Thr	Pro	Ser	Pro	Leu	Gln	Lys	Lys	Val	Thr	Glu	Asp	Leu	
			450				455					460					
15	TCC	AAA	ACC	CTC	CTG	ATG	TAC	ACG	GTG	CCT	GCC	GTC	CAG	GGC	TTC	TTC	1440
	Ser	Lys	Thr	Leu	Leu	MET	Tyr	Thr	Val	Pro	Ala	Val	Gln	Gly	Phe	Phe	
	465					470					475					480	
20	CGT	TCC	ATC	TCC	TTG	TCA	CGA	GGC	AAC	AAC	CTC	CAG	GAT	ACA	CTC	AGA	1488
	Arg	Ser	Ile	Ser	Leu	Ser	Arg	Gly	Asn	Asn	Leu	Gln	Asp	Thr	Leu	Arg	
					485					490					495		
25	GTT	CTC	ACC	TTA	TGG	TTT	GAT	TAT	GGT	CAC	TGG	CCA	GAT	GTC	AAT	GAG	1536
	Val	Leu	Thr	Leu	Trp	Phe	Asp	Tyr	Gly	His	Trp	Pro	Asp	Val	Asn	Glu	
				500				505						510			
30	GCC	TTA	GTG	GAG	GGG	GTG	AAA	GCC	ATC	CAG	ATT	GAT	ACC	TGG	CTA	CAG	1584
	Ala	Leu	Val	Glu	Gly	Val	Lys	Ala	Ile	Gln	Ile	Asp	Thr	Trp	Leu	Gln	
			515					520					525				
35	GTT	ATA	CCT	CAG	CTC	ATT	GCA	AGA	ATT	GAT	ACG	CCC	AGA	CCC	TTG	GTG	1632
	Val	Ile	Pro	Gln	Leu	Ile	Ala	Arg	Ile	Asp	Thr	Pro	Arg	Pro	Leu	Val	
		530					535					540					
40	GGA	CGT	CTC	ATT	CAC	CAG	CTT	CTC	ACA	GAC	ATT	GGT	CGG	TAC	CAC	CCC	1680
	Gly	Arg	Leu	Ile	His	Gln	Leu	Leu	Thr	Asp	Ile	Gly	Arg	Tyr	His	Pro	
	545					550					555					560	
45	CAG	GCC	CTC	ATC	TAC	CCA	CTG	ACA	GTG	GCT	TCT	AAG	TCT	ACC	ACG	ACA	1728
	Gln	Ala	Leu	Ile	Tyr	Pro	Leu	Thr	Val	Ala	Ser	Lys	Ser	Thr	Thr	Thr	
					565				570						575		
50	GCC	CGG	CAC	AAT	GCA	GCC	AAC	AAG	ATT	CTG	AAG	AAC	ATG	TGT	GAG	CAC	1776
	Ala	Arg	His	Asn	Ala	Ala	Asn	Lys	Ile	Leu	Lys	Asn	MET	Cys	Glu	His	
				580				585					590				
45	AGC	AAC	ACC	CTG	GTC	CAG	CAG	GCC	ATG	ATG	GTG	AGC	GAG	GAG	CTG	ATC	1824
	Ser	Asn	Thr	Leu	Val	Gln	Gln	Ala	MET	MET	Val	Ser	Glu	Glu	Leu	Ile	
			595					600					605				
50	CGA	GTG	GCC	ATC	CTC	TGG	CAT	GAG	ATG	TGG	CAT	GAA	GGC	CTG	GAA	GAG	1872
	Arg	Val	Ala	Ile	Leu	Trp	His	Glu	MET	Trp	His	Glu	Gly	Leu	Glu	Glu	
		610					615					620					

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	GCA	TCT	CGT	TTG	TAC	TTT	GGG	GAA	AGG	AAC	GTG	AAA	GGC	ATG	TTT	GAG	1920
	Ala	Ser	Arg	Leu	Tyr	Phe	Gly	Glu	Arg	Asn	Val	Lys	Gly	MET	Phe	Glu	
	625					630					635					640	
5	GTG	CTG	GAG	CCC	TTG	CAT	GCT	ATG	ATG	GAA	CGG	GGC	CCC	CAG	ACT	CTG	1968
	Val	Leu	Glu	Pro	Leu	His	Ala	MET	MET	Glu	Arg	Gly	Pro	Gln	Thr	Leu	
					645					650					655		
10	AAG	GAA	ACA	TCC	TTT	AAT	CAG	GCC	TAT	GGT	CGA	GAT	TTA	ATG	GAG	GCC	2016
	Lys	Glu	Thr	Ser	Phe	Asn	Gln	Ala	Tyr	Gly	Arg	Asp	Leu	MET	Glu	Ala	
				660					665					670			
15	CAA	GAG	TGG	TGC	AGG	AAG	TAC	ATG	AAA	TCA	GGG	AAT	GTC	AAG	GAC	CTC	2064
	Gln	Glu	Trp	Cys	Arg	Lys	Tyr	MET	Lys	Ser	Gly	Asn	Val	Lys	Asp	Leu	
			675					680					685				
20	ACC	CAA	GCC	TGG	GAC	CTC	TAT	TAT	CAT	GTG	TTC	CGA	CGA	ATC	TCA	AAG	2112
	Thr	Gln	Ala	Trp	Asp	Leu	Tyr	Tyr	His	Val	Phe	Arg	Arg	Ile	Ser	Lys	
			690				695					700					
25	CAG	CTG	CCT	CAG	CTC	ACA	TCC	TTA	GAG	CTG	CAA	TAT	GTT	TCC	CCA	AAA	2160
	Gln	Leu	Pro	Gln	Leu	Thr	Ser	Leu	Glu	Leu	Gln	Tyr	Val	Ser	Pro	Lys	
						710					715					720	
30	CTT	CTG	ATG	TGC	CGG	GAC	CTT	GAA	TTG	GCT	GTG	CCA	GGA	ACA	TAT	GAC	2208
	Leu	Leu	MET	Cys	Arg	Asp	Leu	Glu	Leu	Ala	Val	Pro	Gly	Thr	Tyr	Asp	
					725					730					735		
35	CCC	AAC	CAG	CCA	ATC	ATT	CGC	ATT	CAG	TCC	ATA	GCA	CCG	TCT	TTG	CAA	2256
	Pro	Asn	Gln	Pro	Ile	Ile	Arg	Ile	Gln	Ser	Ile	Ala	Pro	Ser	Leu	Gln	
				740					745					750			
40	GTC	ATC	ACA	TCC	AAG	CAG	AGG	CCC	CGG	AAA	TTG	ACA	CTT	ATG	GGC	AGC	2304
	Val	Ile	Thr	Ser	Lys	Gln	Arg	Pro	Arg	Lys	Leu	Thr	Leu	MET	Gly	Ser	
			755					760					765				
45	AAC	GGA	CAT	GAG	TTT	GTT	TTC	CTT	CTA	AAA	GGC	CAT	GAA	GAT	CTG	CGC	2352
	Asn	Gly	His	Glu	Phe	Val	Phe	Leu	Leu	Lys	Gly	His	Glu	Asp	Leu	Arg	
		770					775					780					
50	CAG	GAT	GAG	CGT	GTG	ATG	CAG	CTC	TTC	GGC	CTG	GTT	AAC	ACC	CTT	CTG	2400
	Gln	Asp	Glu	Arg	Val	MET	Gln	Leu	Phe	Gly	Leu	Val	Asn	Thr	Leu	Leu	
						790					795					800	
55	GCC	AAT	GAC	CCA	ACA	TCT	CTT	CGG	AAA	AAC	CTC	AGC	ATC	CAG	AGA	TAC	2448
	Ala	Asn	Asp	Pro	Thr	Ser	Leu	Arg	Lys	Asn	Leu	Ser	Ile	Gln	Arg	Tyr	
					805					810					815		
60	GCT	GTC	ATC	CCT	TTA	TCG	ACC	AAC	TCG	GGC	CTC	ATT	GGC	TGG	GTT	CCC	2496
	Ala	Val	Ile	Pro	Leu	Ser	Thr	Asn	Ser	Gly	Leu	Ile	Gly	Trp	Val	Pro	
				820					825					830			

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	CAC	TGT	GAC	ACA	CTG	CAC	GCC	CTC	ATC	CGG	GAC	TAC	AGG	GAG	AAG	AAG	2544
	His	Cys	Asp	Thr	Leu	His	Ala	Leu	Ile	Arg	Asp	Tyr	Arg	Glu	Lys	Lys	
			835					840					845				
5	AAG	ATC	CTT	CTC	AAC	ATC	GAG	CAT	CGC	ATC	ATG	TTG	CGG	ATG	GCT	CCG	2592
	Lys	Ile	Leu	Leu	Asn	Ile	Glu	His	Arg	Ile	MET	Leu	Arg	MET	Ala	Pro	
		850					855					860					
10	GAC	TAT	GAC	CAC	TTG	ACT	CTG	ATG	CAG	AAG	GTG	GAG	GTG	TTT	GAG	CAT	2640
	Asp	Tyr	Asp	His	Leu	Thr	Leu	MET	Gln	Lys	Val	Glu	Val	Phe	Glu	His	
	865					870					875				880		
15	GCC	GTC	AAT	AAT	ACA	GCT	GGG	GAC	GAC	CTG	GCC	AAG	CTG	CTG	TGG	CTG	2688
	Ala	Val	Asn	Asn	Thr	Ala	Gly	Asp	Asp	Leu	Ala	Lys	Leu	Leu	Trp	Leu	
					885					890					895		
20	AAA	AGC	CCC	AGC	TCC	GAG	GTG	TGG	TTT	GAC	CGA	AGA	ACC	AAT	TAT	ACC	2736
	Lys	Ser	Pro	Ser	Ser	Glu	Val	Trp	Phe	Asp	Arg	Arg	Thr	Asn	Tyr	Thr	
				900					905					910			
25	CGT	TCT	TTA	GCG	GTC	ATG	TCA	ATG	GTT	GGG	TAT	ATT	TTA	GGC	CTG	GGA	2784
	Arg	Ser	Leu	Ala	Val	MET	Ser	MET	Val	Gly	Tyr	Ile	Leu	Gly	Leu	Gly	
			915					920					925				
30	GAT	AGA	CAC	CCA	TCC	AAC	CTG	ATG	CTG	GAC	CGT	CTG	AGT	GGG	AAG	ATC	2832
	Asp	Arg	His	Pro	Ser	Asn	Leu	MET	Leu	Asp	Arg	Leu	Ser	Gly	Lys	Ile	
		930					935					940					
35	CTG	CAC	ATT	GAC	TTT	GGG	GAC	TGC	TTT	GAG	GTT	GCT	ATG	ACC	CGA	GAG	2880
	Leu	His	Ile	Asp	Phe	Gly	Asp	Cys	Phe	Glu	Val	Ala	MET	Thr	Arg	Glu	
	945					950				955						960	
40	AAG	TTT	CCA	GAG	AAG	ATT	CCA	TTT	AGA	CTA	ACA	AGA	ATG	TTG	ACC	AAT	2928
	Lys	Phe	Pro	Glu	Lys	Ile	Pro	Phe	Arg	Leu	Thr	Arg	MET	Leu	Thr	Asn	
				965					970					975			
45	GCT	ATG	GAG	GTT	ACA	GGC	CTG	GAT	GGC	AAC	TAC	AGA	ATC	ACA	TGC	CAC	2976
	Ala	MET	Glu	Val	Thr	Gly	Leu	Asp	Gly	Asn	Tyr	Arg	Ile	Thr	Cys	His	
				980					985					990			
50	ACA	GTG	ATG	GAG	GTG	CTG	CGA	GAG	CAC	AAG	GAC	AGT	GTC	ATG	GCC	GTG	3024
	Thr	Val	MET	Glu	Val	Leu	Arg	Glu	His	Lys	Asp	Ser	Val	MET	Ala	Val	
			995					1000					1005				
55	CTG	GAA	GCC	TTT	GTC	TAT	GAC	CCC	TTG	CTG	AAC	TGG	AGG	CTG	ATG	GAC	3072
	Leu	Glu	Ala	Phe	Val	Tyr	Asp	Pro	Leu	Leu	Asn	Trp	Arg	Leu	MET	Asp	
		1010					1015					1020					
60	ACA	AAT	ACC	AAA	GGC	AAC	AAG	CGA	TCC	CGA	ACG	AGG	ACG	GAT	TCC	TAC	3120
	Thr	Asn	Thr	Lys	Gly	Asn	Lys	Arg	Ser	Arg	Thr	Arg	Thr	Asp	Ser	Tyr	
	1025					1030					1035					1040	

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	TCT GCT GGC CAG TCA GTC GAA ATT TTG GAC GGT GTG GAA CTT GGA GAG	3168
	Ser Ala Gly Gln Ser Val Glu Ile Leu Asp Gly Val Glu Leu Gly Glu	
	1045 1050 1055	
5	CCA GCC CAT AAG AAA ACG GGG ACC ACA GTG CCA GAA TCT ATT CAT TCT	3216
	Pro Ala His Lys Lys Thr Gly Thr Thr Val Pro Glu Ser Ile His Ser	
	1060 1065 1070	
10	TTC ATT GGA GAC GGT TTG GTG AAA CCA GAG GCC CTA AAT AAG AAA GCT	3264
	Phe Ile Gly Asp Gly Leu Val Lys Pro Glu Ala Leu Asn Lys Lys Ala	
	1075 1080 1085	
15	ATC CAG ATT ATT AAC AGG GTT CGA GAT AAG CTC ACT GGT CGG GAC TTC	3312
	Ile Gln Ile Ile Asn Arg Val Arg Asp Lys Leu Thr Gly Arg Asp Phe	
	1090 1095 1100	
	TCT CAT GAT GAC ACT TTG GAT GTT CCA ACG CAA GTT GAG CTG CTC ATC	3360
	Ser His Asp Asp Thr Leu Asp Val Pro Thr Gln Val Glu Leu Leu Ile	
	1105 1110 1115 1120	
20	AAA CAA GCG ACA TCC CAT GAA AAC CTC TGC CAG TGC TAT ATT GGC TGG	3408
	Lys Gln Ala Thr Ser His Glu Asn Leu Cys Gln Cys Tyr Ile Gly Trp	
	1125 1130 1135	
25	TAC CCT TTC TGG TAA	3423
	Tyr Pro Phe Trp	
	1140	